Theophylline

Order information

<table>
<thead>
<tr>
<th>REF</th>
<th>CONTENT</th>
<th>Analyzer(s) on which cobas c pack(s) can be used</th>
</tr>
</thead>
<tbody>
<tr>
<td>04491025</td>
<td>ONLINE TDM Theophylline 100 tests</td>
<td>System-ID 07 6927 4 Roche/Hitachi cobas c 501/502</td>
</tr>
<tr>
<td>03375790</td>
<td>Preciset TDM I calibrators</td>
<td>Codes 691-696</td>
</tr>
<tr>
<td></td>
<td>CAL A-F (1 x 5 mL)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Diluent (1 x 10 mL)</td>
<td></td>
</tr>
<tr>
<td>04521536</td>
<td>TDM Control Set</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Level I (2 x 5 mL)</td>
<td>Code 310</td>
</tr>
<tr>
<td></td>
<td>Level II (2 x 5 mL)</td>
<td>Code 311</td>
</tr>
<tr>
<td></td>
<td>Level III (2 x 5 mL)</td>
<td>Code 312</td>
</tr>
</tbody>
</table>

English

System information
For cobas c 501 analyzer:
THEO2: ACN 415
For cobas c 502 analyzer:
THEO2: ACN 8415

Intended use
In vitro test for the quantitative determination of theophylline in serum and plasma on Roche/Hitachi cobas c systems.

Summary
Theophylline (1,3-dimethylxanthine), a bronchodilator, is widely used to treat patients with asthma, apnea (temporary asphyxia), and other obstructive lung diseases. Monitoring of theophylline concentrations in serum is essential, since individuals can vary in their rates of theophylline clearance,1,2 and severe toxicity has been observed without prior occurrence of minor side effects.3 Moreover, several factors can alter theophylline elimination. Theophylline elimination is slowed in obese patients, patients with hepatic disease, and in those on a high carbohydrate, low protein diet. Premature infants have very low rates of theophylline elimination.4 Conversely, theophylline elimination is more rapid among cigarette smokers.5 In combination with other clinical data, monitoring serum theophylline levels may provide the physician with useful information to aid in adjusting patient dosage to achieve optimal therapeutic effect while avoiding drug toxicity.

Test principle
The assay is based on the kinetic interaction of microparticles in a solution (KIMS). Theophylline antibody is covalently coupled to microparticles and the drug derivative is linked to a macromolecule. The kinetic interaction of microparticles in solutions is induced by binding of drug-conjugate to the antibody on the microparticles and is inhibited by the presence of theophylline in the sample. A competitive reaction takes place between the drug conjugate and theophylline in the serum sample for binding to the theophylline antibody on the microparticles. The resulting kinetic interaction of microparticles is indirectly proportional to the amount of drug present in the sample.

Reagents - working solutions

R1 Theophylline conjugate; piperazine-N,N'-bis (ethanesulfonic acid) (PIES) buffer; pH 7.2; preservative

R2 Anti-theophylline antibody (mouse monoclonal); latex microparticle; 2-[(N-morpholino) propane sulfonic acid (MOPS) buffer; pH 7.5; stabilizer; preservative

R1 is in position B and R2 is in position C.

Precautions and warnings
For in vitro diagnostic use.
Exercise the normal precautions required for handling all laboratory reagents.
Disposal of all waste material should be in accordance with local guidelines.
Safety data sheet available for professional user on request.
For USA: Caution: Federal law restricts this device to sale by or on the order of a physician.

Reagent handling
Ready for use

Carefully invert reagent container several times prior to use to ensure that the reagent components are mixed.

Storage and stability

Storage: -20 °C
Shelf life at 2-8 °C: See expiration date on cobas c pack label

On-board in use and refrigerated on the analyzer: 12 weeks

Do not freeze.

Specimen collection and preparation
For specimen collection and preparation only use suitable tubes or collection containers.
Only the specimens listed below were tested and found acceptable.
Serum: Collect serum using standard sampling tubes Plasma: K₂ or K₃-EDTA, sodium citrate, or sodium, lithium or ammonium heparin plasma.

Stability: 1 week capped at 2-8 °C
60 days capped at -20 °C

The sample types listed were tested with a selection of sample collection tubes that were commercially available at the time of testing, i.e. not all available tubes of all manufacturers were tested. Sample collection systems from various manufacturers may contain differing materials which could affect the test results in some cases. When processing samples in primary tubes (sample collection systems), follow the instructions of the tube manufacturer.
Centrifuge samples containing precipitates before performing the assay. Specimens should not be repeatedly frozen and thawed.
Invert thawed specimens several times prior to testing. Use sampling time varies dependent upon desired measurement of peak or trough values.7

Materials provided
See “Reagents – working solutions” section for reagents.

Materials required (but not provided)
See “Order information” section

General laboratory equipment

Assay
For optimum performance of the assay follow the directions given in this document for the analyzer concerned. Refer to the appropriate operator’s manual for analyzer-specific assay instructions.

The performance of applications not validated by Roche is not warranted and must be defined by the user.

Application for serum and plasma

Deselect Automatic Rerun for these applications in the Utility menu, Application screen, Range tab.

cobas c 501/502 test definition

Assay type 2-Point End

Reaction time /Assay points 10 / 15-49

Wavelength (sub/main) 800/600 nm

Reaction direction Increase
THEO2

Theophylline

Unit
µg/mL

Reagent pipetting
Diluent

R1
97 µL

(H2O)

R2
92 µL

Sample volumes
Sample

Sample dilution

Diluent (NaCl)

Normal
2.0 µL

–

–

Decreased
2.0 µL

–

–

Increased
2.0 µL

–

–

Calibration

Calibrators
S1-6: Preciset TDM I calibrators

Calibration mode
RCM

Calibration frequency
6-point calibration

• after reagent lot change

• every 8 weeks

• as required following quality control procedures

Calibration interval may be extended based on acceptable verification of calibration by the laboratory.

Traceability: This method has been standardized against USP reference standards. The calibrators are prepared to contain known quantities of theophylline in normal human serum.

Quality control

For quality control, use control materials as listed in the "Order information" section.

In addition, other suitable control material can be used.

The control intervals and limits should be adapted to each laboratory's individual requirements. Values obtained should fall within the defined limits. Each laboratory should establish corrective measures to be taken if values fall outside the defined limits.

Follow the applicable government regulations and local guidelines for quality control.

Calculation

Roche/Hitachi cobas c systems automatically calculate the analyte concentration of each sample.

Conversion factor: 1 µg/mL x 5.55 = µmol/L

Limitations - interference

Criterions: Recovery within ± 10 % of initial value at theophylline levels of approximately 5 and 15 µg/mL (27.8 and 83.3 µmol/L).

Serum/Plasma

Icterus: No significant interference up to an I index of 50 for conjugated bilirubin and unconjugated bilirubin (approximate conjugated and unconjugated bilirubin concentration: 50 mg/dL or 855 µmol/L).

Hemolysis: No significant interference up to an H index of 1000 (approximate hemoglobin concentration: 1000 mg/dL or 621 µmol/L).

Lipemia (Intralipid): No significant interference up to an L index of 300.

There is poor correlation between the L index (corresponds to turbidity) and triglycerides concentration.

No significant interference from triglycerides up to 1000 mg/dL (11.3 mmol/L).

Rheumatoid factors: No significant interference from rheumatoid factors up to 100 IU/mL.

Total protein: No interference from total protein up to 12 g/dL.

Theobromine: No significant interference up to 49 µg/mL theobromine.

Concentrations above this toxic level may result in negative bias of > 10 %.

In very rare cases, gammapathy, in particular type IgM (Waldenström's macroglobulinemia), may cause unreliable results.11

For diagnostic purposes, the results should always be assessed in conjunction with the patient's medical history, clinical examination and other findings.

ACTION REQUIRED

Special Wash Programming: The use of special wash steps is mandatory when certain test combinations are run together on Roche/Hitachi cobas c systems. The latest version of the carry-over evasion list can be found with the Roche/Hitachi cobas c 502 Method Sheets. For further instructions refer to the operator's manual. cobas c 502 analyzer: All special wash programming necessary for avoiding carry-over is available via the cobas link, manual input is not required.

Where required, special wash/carry-over evasion programming must be implemented prior to reporting results with this test.

Limits and ranges

Measuring range
0.8-40.0 µg/mL (4.4-222 µmol/L)

Manually dilute samples above the measuring range 1 + 1 with the Preciset TDM I diluent (0 µg/mL) and reassy. Multiply the result by 2 to obtain the specimen value.

Lower limits of measurement

Lower detection limit of the test
0.8 µg/mL (4.4 µmol/L)

The lower detection limit represents the lowest measurable analyte level that can be distinguished from zero. It is calculated as the value lying 2 standard deviations above that of the 0 µg/mL calibrator (standard 1 + 2 SD, repeatability, n = 21).

Expected values

Various methodologies have been used to evaluate theophylline preparations and routes of administration,12 to study pharmacokinetics of the drug,13 and to define the relationship between serum concentration and the drug's therapeutic and toxic effects.14 For most patients, the range of 10 to 20 µg/mL (55.5 to 111 µmol/L) suppresses chronic asthmatic symptoms.10,16,17,18 Wide discrepancies between drug dosage and serum concentrations were observed among patients.12,13 A major factor accounting for the variability is individual variation in the rate of theophylline metabolism and elimination.

Each laboratory should investigate the transferability of the expected values to its own patient population and if necessary determine its own reference ranges.

Specific performance data

Representative performance data on the analyzers are given below.

Results obtained in individual laboratories may differ.

Precision

Precision was determined using human samples and controls in a modified NCCLS EP5-T2 protocol (repeatability n = 63, intermediate precision n = 63). The following results were obtained on a Roche/Hitachi cobas c 501 analyzer.

Serum/Plasma

<table>
<thead>
<tr>
<th>Repeatability</th>
<th>Mean µg/mL</th>
<th>Mean µmol/L</th>
<th>SD µg/mL</th>
<th>SD µmol/L</th>
<th>CV %</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control 1</td>
<td>4.25</td>
<td>23.6</td>
<td>0.07</td>
<td>0.4</td>
<td>1.7</td>
</tr>
<tr>
<td>Control 2</td>
<td>14.3</td>
<td>79.4</td>
<td>0.2</td>
<td>1.1</td>
<td>1.3</td>
</tr>
<tr>
<td>Control 3</td>
<td>34.1</td>
<td>189</td>
<td>0.4</td>
<td>2</td>
<td>1.2</td>
</tr>
<tr>
<td>HS 1</td>
<td>5.78</td>
<td>32.1</td>
<td>0.08</td>
<td>0.4</td>
<td>1.4</td>
</tr>
<tr>
<td>HS 2</td>
<td>20.0</td>
<td>111</td>
<td>0.3</td>
<td>2</td>
<td>1.4</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Intermediate precision</th>
<th>Mean µg/mL</th>
<th>Mean µmol/L</th>
<th>SD µg/mL</th>
<th>SD µmol/L</th>
<th>CV %</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control 1</td>
<td>4.25</td>
<td>23.6</td>
<td>0.12</td>
<td>0.7</td>
<td>2.8</td>
</tr>
<tr>
<td>Control 2</td>
<td>14.3</td>
<td>79.4</td>
<td>0.2</td>
<td>1.1</td>
<td>1.7</td>
</tr>
<tr>
<td>Control 3</td>
<td>34.1</td>
<td>189</td>
<td>0.6</td>
<td>3</td>
<td>1.9</td>
</tr>
<tr>
<td>HS 1</td>
<td>5.78</td>
<td>32.1</td>
<td>0.12</td>
<td>0.7</td>
<td>2.1</td>
</tr>
</tbody>
</table>
Theo2

Theophylline

<table>
<thead>
<tr>
<th>Method comparison</th>
<th>Serum/plasma</th>
</tr>
</thead>
</table>
| Theophylline values for human serum and plasma samples obtained on a Roche/Hitachi cobas c 501 analyzer (y) were compared with those determined using the corresponding reagent on a Roche/Hitachi 917 analyzer (x) and on a COBAS INTEGRA 800 analyzer (x).

**Roche/Hitachi 917 analyzer**
- Sample size (n) = 72
- Linear regression
- y = 0.975x + 0.136 µg/mL
- \( \tau = 0.981 \)

**COBAS INTEGRA 800 analyzer**
- Sample size (n) = 72
- Linear regression
- y = 1.017x + 0.091 µg/mL
- \( \tau = 0.981 \)

The sample concentrations were between 3.98 and 39.0 µg/mL (22.1 and 217 µmol/L).

**Analytical specificity**
- The following compounds were tested for cross-reactivity.

<table>
<thead>
<tr>
<th>Compound</th>
<th>Concentration Tested (µg/mL)</th>
<th>% Cross-reactivity</th>
</tr>
</thead>
<tbody>
<tr>
<td>Aminophylline</td>
<td>15</td>
<td>79.6</td>
</tr>
<tr>
<td>8-Chlorotheophylline</td>
<td>200</td>
<td>5.97</td>
</tr>
<tr>
<td>1,7-Dimethylxanthine</td>
<td>150</td>
<td>5.24</td>
</tr>
<tr>
<td>3-Methylxanthine</td>
<td>150</td>
<td>2.73</td>
</tr>
<tr>
<td>Ephedrine</td>
<td>12</td>
<td>1.00</td>
</tr>
<tr>
<td>Acetaminophen</td>
<td>200</td>
<td>&lt; 1.0</td>
</tr>
<tr>
<td>Allopurinol</td>
<td>50</td>
<td>&lt; 1.0</td>
</tr>
<tr>
<td>Caffeine</td>
<td>150</td>
<td>&lt; 1.0</td>
</tr>
<tr>
<td>Dihydroxypropyl theophylline</td>
<td>200</td>
<td>&lt; 1.0</td>
</tr>
<tr>
<td>Diphenhydramine</td>
<td>10</td>
<td>&lt; 1.0</td>
</tr>
<tr>
<td>Epinephrine</td>
<td>16</td>
<td>&lt; 1.0</td>
</tr>
<tr>
<td>( \beta )-Hydroxyethyl theophylline</td>
<td>200</td>
<td>&lt; 1.0</td>
</tr>
<tr>
<td>7-( \beta )-Hydroxypropyl theophylline</td>
<td>200</td>
<td>&lt; 1.0</td>
</tr>
<tr>
<td>Hypoxanthine</td>
<td>150</td>
<td>&lt; 1.0</td>
</tr>
<tr>
<td>Isoproterenol</td>
<td>50</td>
<td>&lt; 1.0</td>
</tr>
<tr>
<td>1-Methyluric acid</td>
<td>400</td>
<td>&lt; 1.0</td>
</tr>
<tr>
<td>Phenobarbital</td>
<td>200</td>
<td>&lt; 1.0</td>
</tr>
<tr>
<td>Phenylbutazone</td>
<td>400</td>
<td>&lt; 1.0</td>
</tr>
<tr>
<td>Uric acid</td>
<td>210</td>
<td>&lt; 1.0</td>
</tr>
<tr>
<td>1,3-Dimethyluric acid</td>
<td>700</td>
<td>&lt; 0.1</td>
</tr>
<tr>
<td>Phenyltoxin</td>
<td>200</td>
<td>&lt; 0.1</td>
</tr>
</tbody>
</table>

Tests were performed on 15 drugs. No significant interference with the assay was found.

<table>
<thead>
<tr>
<th>Compound</th>
<th>Cross-reactivity</th>
</tr>
</thead>
<tbody>
<tr>
<td>Acetaminophen</td>
<td>Doxycycline (Tetracycline)</td>
</tr>
<tr>
<td>Acetyl cysteine</td>
<td>Ibuprofen</td>
</tr>
<tr>
<td>Acetylsalicylic acid</td>
<td>Levodopa</td>
</tr>
<tr>
<td>Ampicillin-Na</td>
<td>Methyldopa + 1.5 H₂O</td>
</tr>
<tr>
<td>Ascorbic acid</td>
<td>Metronidazole</td>
</tr>
<tr>
<td>Ca-Dobesilate</td>
<td>Phenytoin</td>
</tr>
<tr>
<td>Cefoxitin</td>
<td>Rifampicin</td>
</tr>
<tr>
<td>Cyclosporine</td>
<td></td>
</tr>
</tbody>
</table>

References

A point (period/stop) is always used in this Method Sheet as the decimal separator to mark the border between the integral and the fractional parts of a decimal numeral. Separators for thousands are not used.

Symbols
Roche Diagnostics uses the following symbols and signs in addition to those listed in the ISO 15223-1 standard (for USA: see https://usadiagnostics.roche.com for definition of symbols used):
Voluem after reconstitution or mixing

Global Trade Item Number

FOR US CUSTOMERS ONLY: LIMITED WARRANTY
Roche Diagnostics warrants that this product will meet the specifications stated in the labeling when used in accordance with such labeling and will be free from defects in material and workmanship until the expiration date printed on the label. THIS LIMITED WARRANTY IS IN LIEU OF ANY OTHER WARRANTY, EXPRESS OR IMPLIED, INCLUDING ANY IMPLIED WARRANTY OF MERCHANTABILITY OR FITNESS FOR PARTICULAR PURPOSE. IN NO EVENT SHALL ROCHE DIAGNOSTICS BE LIABLE FOR INCIDENTAL, INDIRECT, SPECIAL OR CONSEQUENTIAL DAMAGES.

COBAS, COBAS C, COBAS INTEGRA, ONLINE TDM and PRECISET are trademarks of Roche.
All other product names and trademarks are the property of their respective owners.
Additions, deletions or changes are indicated by a change bar in the margin.
© 2017, Roche Diagnostics