

Albumin Gen.2

Order information



REF	Ţ <u>i</u>	CONTENT			Analyzer(s) on which cobas c pack(s) can be used
08056692190	08056692500	Albumin Gen.2 (750 tests)		System-ID 2009 001	cobas c 303, cobas c 503, cobas c 703
Materials required (but not provided):				
10759350190	Calibrator f.a.s. (12 x 3 mL)		Code 20401		
05117003190	PreciControl ClinChem Multi 1 (20 x 5 mL)		Code 203	91	
05947626190	PreciControl ClinC	hem Multi 1 (4 x 5 mL)	Code 203	91	

English

System information ALB2-G: ACN 20090

Intended use

05117216190

05947774190

08063494190

In vitro test for the quantitative determination of albumin in human serum and plasma on **cobas c** systems.

Diluent NaCl 9 % (123 mL)

PreciControl ClinChem Multi 2 (20 x 5 mL)

PreciControl ClinChem Multi 2 (4 x 5 mL)

Summary

Albumin measurement in human serum and plasma with this assay can be used to aid in the assessment of hyperalbuminemia (seen only in case of dehydration) or hypoalbuminemia (seen in a multitude of clinical conditions such as inflammation, liver diseases, inflammatory disease of the intestinal tract, tissue damage like burns, nephrotic disease or neoplastic disease).

Albumin is a carbohydrate-free protein, which constitutes 55-65 % of total plasma protein. It maintains plasma oncotic pressure, is involved in the transport and storage of a wide variety of ligands and is a source of endocenous amino acids.¹

In serum and plasma, hyperalbuminemia is of little diagnostic significance except in dehydration. Hypoalbuminemia instead is very common in many diseases and is caused by several factors: impaired synthesis, either primary as a result of a liver disease or secondary due to diminished protein intake; increased catabolism because of tissue damage (severe burns) or inflammation; malabsorption of amino acids or increased gastrointestinal loss (inflammatory bowel disease such as Crohn's disease and ulcerative colitis); proteinuria due to nephrotic syndrome; negative protein and energy balance due to neoplastic disease(s).^{2,3,4}

In severe cases of hypoalbuminemia, plasma albumin levels are below 25 g/L (380 μ mol/L). The low plasma oncotic pressure allows water to move out of the blood capillaries into the tissues (edema). Albumin measurements also allow monitoring of the patient's response to nutritional support and are a useful test of liver function. 1,5,6

Test principle7

Colorimetric assay

At a pH value of 4.1, albumin displays a sufficiently cationic character to be able to bind with bromcresol green (BCG), an anionic dye, to form a blue-green complex.

The color intensity of the blue-green color is directly proportional to the albumin concentration in the sample and is measured photometrically.

Reagents - working solutions

R1 Citrate buffer: 95 mmol/L, pH 4.1; preservatives, stabilizers

R3 Citrate buffer: 95 mmol/L, pH 4.1; bromcresol green: 0.66 mmol/L; preservatives, stabilizers

R1 is in position B and R3 is in position C.

Precautions and warnings

For in vitro diagnostic use for health care professionals. Exercise the normal precautions required for handling all laboratory reagents.

Infectious or microbial waste:

Warning: handle waste as potentially biohazardous material. Dispose of waste according to accepted laboratory instructions and procedures.

Environmental hazards:

Apply all relevant local disposal regulations to determine the safe disposal.

Safety data sheet available for professional user on request.

Reagent handling

Ready for use

Code 20392

Code 20392 System-ID 2906 001

Storage and stability

Shelf life at 15-25 °C: See expiration date on **cobas c** pack label.

On-board in use and refrigerated on the 26 weeks

analyzer:

Specimen collection and preparation

For specimen collection and preparation only use suitable tubes or collection containers.

Only the specimens listed below were tested and found acceptable. Serum.

Plasma: Li-heparin and K2-EDTA plasma

Do not use fluoride plasma.

The sample types listed were tested with a selection of sample collection tubes that were commercially available at the time of testing, i.e. not all available tubes of all manufacturers were tested. Sample collection systems from various manufacturers may contain differing materials which could affect the test results in some cases. When processing samples in primary tubes (sample collection systems), follow the instructions of the tube

Centrifuge samples containing precipitates before performing the assay. See the limitations and interferences section for details about possible sample interferences.

Stability:⁸ 2.5 months at 20-25 °C

5 months at 4-8 °C

4 months at -20 °C (± 5 °C)

Freeze only once.

Materials provided

See "Reagents – working solutions" section for reagents.

Materials required (but not provided)

See "Order information" section

General laboratory equipment

Assay

For optimum performance of the assay follow the directions given in this document for the analyzer concerned. Refer to the appropriate operator's manual for analyzer-specific assay instructions.

The performance of applications not validated by Roche is not warranted and must be defined by the user.





Application for serum and plasma

Test definition

Reporting time 10 min
Wavelength (sub/main) 505/570 nm

Reagent pipetting Diluent (H₂O)

R1 80 μL – R3 16 μL 24 μL

Sample volumes Sample Sample dilution

For further information about the assay test definitions refer to the application parameters setting screen of the corresponding analyzer and assay.

Calibration

Calibrators S1: H₂O

S2: C.f.a.s.

Calibration mode Linear

Calibration frequency Automatic full calibration

- after reagent lot change

Full calibration

- every 4 weeks on-board

- as required following quality control

procedures

Calibration interval may be extended based on acceptable verification of calibration by the laboratory.

Traceability: This method has been standardized against the reference preparation of the IRMM (Institute for Reference Materials and Measurements) BCR470/CRM470 (RPPHS - Reference Preparation for Proteins in Human Serum).9

Quality control

For quality control, use control materials as listed in the "Order information" section. In addition, other suitable control material can be used.

The control intervals and limits should be adapted to each laboratory's individual requirements. It is recommended to perform quality control always after lot calibration and subsequently at least every 26 weeks.

Values obtained should fall within the defined limits. Each laboratory should establish corrective measures to be taken if values fall outside the defined limits

Follow the applicable government regulations and local guidelines for quality control.

Calculation

cobas c systems automatically calculate the analyte concentration of each sample in the unit g/L (μ mol/L, g/dL).

Conversion factors: $g/L \times 15.2 = \mu mol/L$

 $g/L \times 0.1 = g/dL$

Limitations - interference

Criterion: Recovery within \pm 3.5 g/L of initial values of samples \leq 35 g/L and within \pm 10 % for samples > 35 g/L.

Icterus: ¹⁰ No significant interference up to an I index of 60 for conjugated and unconjugated bilirubin (approximate conjugated and unconjugated bilirubin concentration: 1026 µmol/L or 60 mg/dL).

Hemolysis: 10 No significant interference up to an H index of 1000 (approximate hemoglobin concentration: 621 μ mol/L or 1000 mg/dL).

Lipemia (Intralipid): ¹⁰ No significant interference up to an L index of 550. There is poor correlation between the L index (corresponds to turbidity) and triglycerides concentration.

Drugs: No interference was found at the rapeutic concentrations using common drug panels. $^{11,12}\,$

In very rare cases, gammopathy, in particular type IgM (Waldenström's macroglobulinemia), may cause unreliable results.¹³

For diagnostic purposes, the results should always be assessed in conjunction with the patient's medical history, clinical examination and other findings.

Colorimetric methods used for the determination of Albumin may lead to falsely elevated test results in patients suffering from renal failure or insufficiency due to interference with other proteins. Immunoturbidimetric methods are less affected.

ACTION REQUIRED

Special Wash Programming: The use of special wash steps is mandatory when certain test combinations are run together on **cobas c** systems. All special wash programming necessary for avoiding carry-over is available via the **cobas** link. The latest version of the carry-over evasion list can be found with the NaOHD/SMS/SCCS Method Sheet. For further instructions, refer to the operator's manual.

Limits and ranges

Measuring range

2-60 g/L (30.4-912 µmol/L)

Determine samples having higher concentrations via the rerun function. Dilution of samples via the rerun function is a 1:3 dilution. Results from samples diluted using the rerun function are automatically multiplied by a factor of 3.

Lower limits of measurement

Limit of Blank, Limit of Detection and Limit of Quantitation

 $\begin{array}{ll} \mbox{Limit of Blank} & = 2 \mbox{ g/L } (30.4 \mbox{ } \mu \mbox{mol/L}) \\ \mbox{Limit of Detection} & = 2 \mbox{ g/L } (30.4 \mbox{ } \mu \mbox{mol/L}) \\ \mbox{Limit of Quantitation} & = 3 \mbox{ g/L } (45.6 \mbox{ } \mu \mbox{mol/L}) \end{array}$

The Limit of Blank, Limit of Detection and Limit of Quantitation were determined in accordance with the CLSI (Clinical and Laboratory Standards Institute) EP17-A2 requirements.

The Limit of Blank is the 95^{th} percentile value from $n \ge 60$ measurements of analyte-free samples over several independent series. The Limit of Blank corresponds to the concentration below which analyte-free samples are found with a probability of 95%.

The Limit of Detection is determined based on the Limit of Blank and the standard deviation of low concentration samples.

The Limit of Detection corresponds to the lowest analyte concentration which can be detected (value above the Limit of Blank with a probability of 95%)

The Limit of Quantitation is the lowest analyte concentration that can be reproducibly measured with a total error of 20 %. It has been determined using low concentration albumin samples.

Expected values

g/L

Reference range study14

Adults 39.7-49.4 g/L

Consensus values¹⁵

Adults 35-52 g/L

Reference intervals according to Tietz¹⁶

Newborn

0-4 days 28-44 g/L

Children

4 days-14 years 38-54 g/L 14-18 years 32-45 g/L

umol/L*

* calculated by unit conversion factor

Reference range study¹⁴

Adults 603-751 µmol/L





Consensus values¹⁵

Adults 532-790 µmol/L

Reference intervals according to Tietz¹⁶

Newborn

0-4 days 426-669 μmol/L

Children

4 days-14 years 578-821 μmol/L 14-18 years 486-684 μmol/L

Each laboratory should investigate the transferability of the expected values to its own patient population and if necessary determine its own reference ranges.

Specific performance data

Representative performance data on the analyzers are given below. These data represent the performance of the analytical procedure itself.

Results obtained in individual laboratories may differ due to heterogenous sample materials, aging of analyzer components and mixture of reagents running on the analyzer.

Precision

Precision was determined using human samples and controls in accordance with the CLSI (Clinical and Laboratory Standards Institute) EP05-A3 requirements with repeatability (n = 84) and intermediate precision (2 aliquots per run, 2 runs per day, 21 days). Results for repeatability and intermediate precision were obtained on the **cobas c** 503 analyzer.

Repeatability	Mean g/L	SD g/L	CV %
PCCC1a)	33.9	0.270	0.8
PCCC2 ^{b)}	47.2	0.223	0.5
Human serum 1	52.3	0.252	0.5
Human serum 2	16.0	0.245	1.5
Human serum 3	32.7	0.280	0.9
Human serum 4	45.6	0.253	0.6
Human serum 5	49.5	0.258	0.5
Intermediate precision	Mean g/L	SD g/L	CV %
Intermediate precision PCCC1a)			•
,	g/L	g/L	%
PCCC1a)	g/L 33.9	g/L 0.865	% 2.6
PCCC1 ^{a)} PCCC2 ^{b)}	g/L 33.9 48.9	g/L 0.865 0.878	% 2.6 1.8
PCCC1 ^{a)} PCCC2 ^{b)} Human serum 1	g/L 33.9 48.9 52.3	g/L 0.865 0.878 0.656	% 2.6 1.8 1.3
PCCC1 ^{a)} PCCC2 ^{b)} Human serum 1 Human serum 2	g/L 33.9 48.9 52.3 16.0	g/L 0.865 0.878 0.656 1.00	2.6 1.8 1.3 6.2

a) PreciControl ClinChem Multi 1

The data obtained on **cobas c** 503 analyzer(s) are representative for **cobas c** 303 analyzer(s) and **cobas c** 703 analyzer(s).

Method comparison

Albumin values for human serum and plasma samples obtained on a **cobas c** 503 analyzer (y) were compared with those determined using the corresponding reagent on a **cobas c** 501 analyzer (x).

Sample size (n) = 142

Passing/Bablok¹⁷ Linear regression y = 0.987x + 1.75 g/L y = 0.999x + 1.26 g/L t = 0.851 t = 0.992

The sample concentrations were between 2.60 and 57.7 g/L.

Albumin values for human serum and plasma samples obtained on a **cobas c** 303 analyzer (y) were compared with those determined using the corresponding reagent on a **cobas c** 501 analyzer (x).

Sample size (n) = 72

 $\begin{aligned} & Passing/Bablok^{17} & Linear\ regression \\ & y = 1.004x + 0.719\ g/L & y = 1.001x + 0.852\ g/L \end{aligned}$

T = 0.922 r = 0.998

The sample concentrations were between 2.84 and 57.2 g/L.

Albumin values for human serum and plasma samples obtained on a **cobas c** 703 analyzer (y) were compared with those determined using the corresponding reagent on a **cobas c** 503 analyzer (x).

Sample size (n) = 75

Passing/Bablok¹⁷ Linear regression y = 1.005x - 0.450 g/L y = 1.003x - 0.376 g/Lz = 0.971 z = 0.999

The sample concentrations were between 3.97 and 58.8 g/L.

References

- Dietzen DJ, Willrich MAV. Amino acids, peptides, and proteins. In: Rifai N, Chiu RWK, Young I, Burnham CAD, Wittwer CT, editors. Tietz Textbook of Laboratory Medicine, Saunders Elsevier, Philadelphia, 7th edition, 2023, chapter 31, p. 348-349.e42.
- 2 Levitt DG, Levitt MD. Human serum albumin homeostasis: a new look at the roles of synthesis, catabolism, renal and gastrointestinal excretion, and the clinical value of serum albumin measurements. Int J Gen Med 2016 Jul 15;9:229-255. doi: 10.2147/IJGM.S102819.
- 3 Gatta A, Verardo A, Bolognesi M. Hypoalbuminemia. Intern Emerg Med 2012 Oct;7 Suppl 3:S193-199.doi: 10-1007/s11739-012-0802-0.
- Furfaro F, Bezzio C, Maconi G. Protein-losing enteropathy in inflammatory bowel diseases. Minerva Gastroenterol Dietol 2015 Dec;61(4):261-265. Epub 2015 Oct 7.
- 5 Cederholm T, Barazzoni R, Austin P, et al. ESPEN guidelines on definitions and terminology of clinical nutrition. Clin Nutr 2017 Feb;36(1):49-64. doi: 10.1016/j.clnu.2016.09.004.
- 6 Johnson PJ, Berhane S, Kagebayashi C, et al. Assessment of liver function in patients with hepatocellular carcinoma: a new evidencebased approach-the ALBI grade. J Clin Oncol 2015 Feb 20;33(6):550-558. doi: 10.1200/JCO.2014.57.9151.
- 7 Doumas BT, Watson WA, Biggs HG. Albumin standards and the measurement of serum albumin with bromcresol green. Clin Chim Acta 1971;31:87-96.
- 8 Use of Anticoagulants in Diagnostic Laboratory Investigations. WHO Publication WHO/DIL/LAB/99.1 Rev. 2: Jan 2002.
- 9 Baudner S, Bienvenu J, Blirup-Jensen S, et al. The certification of a matrix reference material for immunochemical measurement of 14 human serum proteins CRM470. Report EUR 15243 EN 1993;1-186.
- 10 Glick MR, Ryder KW, Jackson SA. Graphical Comparisons of Interferences in Clinical Chemistry Instrumentation. Clin Chem 1986;32:470-475.
- 11 Breuer J. Report on the Symposium "Drug effects in Clinical Chemistry Methods". Eur J Clin Chem Clin Biochem 1996;34:385-386.
- 12 Sonntag O, Scholer A. Drug interference in clinical chemistry: recommendation of drugs and their concentrations to be used in drug interference studies. Ann Clin Biochem 2001;38:376-385.
- 13 Bakker AJ, Mücke M. Gammopathy interference in clinical chemistry assays: mechanisms, detection and prevention. Clin Chem Lab Med 2007;45(9):1240-1243.
- 14 Junge W, Bossert-Reuther S, Klein G, et al. Reference Range Study for Serum Albumin using different methods. Clin Chem Lab Med (June 2007 Poster EUROMEDLAB) 2007;45 Suppl:194.

b) PreciControl ClinChem Multi 2





- 15 Dati F, Schumann G, Thomas L, et al. Consensus of a group of professional societies and diagnostic companies on guidelines for interim reference ranges for 14 proteins in serum based on the standardization against the IFCC/BCR/CAP reference material (CRM 470). Eur J Clin Chem Clin Biochem 1996;34:517-520.
- 16 Burtis CA, Ashwood ER, Bruns DE (eds.). Tietz Textbook of Clinical Chemistry and Molecular Diagnostics. 4th ed. St Louis, Missouri; Elsevier Saunders 2006;549.
- 17 Bablok W, Passing H, Bender R, et al. A general regression procedure for method transformation. Application of linear regression procedures for method comparison studies in clinical chemistry, Part III. J Clin Chem Clin Biochem 1988 Nov;26(11):783-790.

A point (period/stop) is always used in this Method Sheet as the decimal separator to mark the border between the integral and the fractional parts of a decimal numeral. Separators for thousands are not used.

Any serious incident that has occurred in relation to the device shall be reported to the manufacturer and the competent authority of the Member State in which the user and/or the patient is established.

Roche Diagnostics uses the following symbols and signs in addition to those listed in the ISO 15223-1 standard:



Contents of kit

Volume for reconstitution

Global Trade Item Number

Rx only

For USA: Caution: Federal law restricts this device to sale by or on the order of a physician.

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