



Aspartate Aminotransferase acc. to IFCC with pyridoxal phosphate activation

Order information

REF	CONTENT		Analyzer(s) on which cobas c pack(s) can be used
08056838190	Aspartate Aminotransferase acc. to IFCC (500 tests)	System-ID 2022 001	cobas c 303, cobas c 503
Materials required	d (but not provided):		•
10759350190	Calibrator f.a.s. (12 x 3 mL)	Code 20401	
05117003190	PreciControl ClinChem Multi 1 (20 x 5 mL)	Code 20391	
05947626190	PreciControl ClinChem Multi 1 (4 x 5 mL)	Code 20391	
05117216190	PreciControl ClinChem Multi 2 (20 x 5 mL)	Code 20392	
05947774190	PreciControl ClinChem Multi 2 (4 x 5 mL)	Code 20392	
08063494190	Diluent NaCl 9 % (123 mL)	System-ID 2906 001	
08062986190	Pyridoxal phosphate (950 tests)	System-ID 2012 001	

English

System information ASTP: ACN 20220

Intended use

In vitro test for the quantitative determination of aspartate aminotransferase (AST) with pyridoxal phosphate activation in human serum and plasma on ${\bf cobas} \ {\bf c}$ systems.

Summary^{1,2}

The enzyme aspartate aminotransferase (AST) is widely distributed in tissue, principally hepatic, cardiac, muscle, and kidney. Elevated serum levels are found in diseases involving these tissues. Hepatobiliary diseases, such as cirrhosis, metastatic carcinoma, and viral hepatitis also increase serum AST levels. Following myocardial infarction, serum AST is elevated and reaches a peak 2 days after onset.

2 isoenzymes of AST have been detected, cytoplasmic and mitochondrial. Only the cytoplasmic isoenzyme occurs in normal serum, while the mitochondrial, together with the cytoplasmic isoenzyme, has been detected in the serum of patients with coronary and hepatobiliary disease.

The addition of pyridoxal phosphate to the assay causes an increase in aminotransferase activity. The activation is higher for AST than for ALT. Pyridoxal phosphate activation prevents falsely low aminotransferase activity in patient samples with insufficient endogenous pyridoxal phosphate (vitamin B_{θ} deficiency).

Test principle

This assay follows the recommendations of the IFCC, but was optimized for performance and stability. $^{3.4}$

AST in the sample catalyzes the transfer of an amino group between L-aspartate and 2-oxoglutarate to form oxaloacetate and L-glutamate. The oxaloacetate then reacts with NADH, in the presence of malate dehydrogenase (MDH), to form NAD+. Pyridoxal phosphate serves as a coenzyme in the amino transfer reaction. It ensures full enzyme activation.

The rate of the NADH oxidation is directly proportional to the catalytic AST activity. It is determined by measuring the decrease in absorbance.

Reagents - working solutions

Aspartate Aminotransferase acc. to IFCC (ASTL)

R1 TRIS buffer: 264 mmol/L, pH 7.8 (37 °C); L-aspartate: 792 mmol/L; MDH (microorganism): ≥ 24 μkat/L; LDH (microorganisms): ≥ 48 μkat/L; albumin (bovine): 0.25 %; preservative

R3 NADH: ≥ 1.7 mmol/L; 2-oxoglutarate: 94 mmol/L; preservative

R1 is in position B and R3 is in position C.

Pyridoxal phosphate (PYP, Cat. No 08062986190)

R2 Pyridoxal phosphate: 730 μmol/L; additives; preservative

R2 is in position B.

Precautions and warnings

For in vitro diagnostic use for health care professionals. Exercise the normal precautions required for handling all laboratory reagents.

Infectious or microbial waste:

Warning: handle waste as potentially biohazardous material. Dispose of waste according to accepted laboratory instructions and procedures.

Environmental hazards:

Apply all relevant local disposal regulations to determine the safe disposal. Safety data sheet available for professional user on request.

Reagent handling

Ready for use

Storage and stability

Shelf life at 2-8 °C: See expiration date on **cobas c** pack label.

On-board in use and refrigerated on the

12 weeks

analyzer:

Specimen collection and preparation

For specimen collection and preparation only use suitable tubes or collection containers.

Only the specimens listed below were tested and found acceptable. Serum.

Plasma: Li-heparin and K2-EDTA plasma

The sample types listed were tested with a selection of sample collection tubes that were commercially available at the time of testing, i.e. not all available tubes of all manufacturers were tested. Sample collection systems from various manufacturers may contain differing materials which could affect the test results in some cases. When processing samples in primary tubes (sample collection systems), follow the instructions of the tube manufacturer.

Centrifuge samples containing precipitates before performing the assay. See the limitations and interferences section for details about possible sample interferences.

Stability:⁵ 4 days at 20-25 °C

7 days at 4-8 °C 3 months at -20 °C

Materials provided

See "Reagents - working solutions" section for reagents.

Materials required (but not provided)

See "Order information" section General laboratory equipment

Assay

For optimum performance of the assay follow the directions given in this document for the analyzer concerned. Refer to the appropriate operator's manual for analyzer-specific assay instructions.





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The performance of applications not validated by Roche is not warranted and must be defined by the user.

Application for serum and plasma

Test definition

Reporting time 10 min 700/340 nm Wavelength (sub/main) Reagent pipetting Diluent (H₂O) R1 30 uL 38 µL R2 15 µL R3 15 µL 15 µL Sample volumes Sample Sample dilution Diluent (NaCl) Sample Normal $6.8 \mu L$

For further information about the assay test definitions refer to the application parameters setting screen of the corresponding analyzer and assay.

6.8 µL

6.8 µL

Calibration

Decreased

Increased

Calibrators S1: H_2O S2: C.f.a.s.

Calibration mode Linear

Calibration frequency Automatic full calibration

- after reagent lot change

Full calibration

- as required following quality control

10 µL

90 µL

procedures

Calibration interval may be extended based on acceptable verification of calibration by the laboratory.

Traceability: This method has been standardized against the original IFCC formulation using calibrated pipettes together with a manual photometer providing absolute values and the substrate-specific absorptivity, ϵ .⁶

Quality control

For quality control, use control materials as listed in the "Order information" section. In addition, other suitable control material can be used.

The control intervals and limits should be adapted to each laboratory's individual requirements. It is recommended to perform quality control always after lot calibration and subsequently at least every 12 weeks. Values obtained should fall within the defined limits. Each laboratory should establish corrective measures to be taken if values fall outside the defined limits.

Follow the applicable government regulations and local guidelines for quality control.

Calculation

cobas c systems automatically calculate the analyte activity of each sample in the unit U/L ($\mu kat/L$).

Conversion factor: $U/L \times 0.0167 = \mu kat/L$

Limitations - interference

Criterion: Recovery within \pm 10 % of initial value at an AST activity of 35 U/L.

Icterus:⁷ No significant interference up to an I index of 60 for conjugated and unconjugated bilirubin (approximate conjugated and unconjugated bilirubin concentration: 1026 µmol/L or 60 mg/dL).

Hemolysis: No significant interference up to an H index of 20 (approximate hemoglobin concentration: $12.4 \, \mu mol/L$ or $20 \, mg/dL$).

Contamination with erythrocytes will elevate results, because the analyte level in erythrocytes is higher than in normal sera. The level of interference may be variable depending on the content of analyte in the lysed erythrocytes.

Lipemia (Intralipid): No significant interference up to an L index of 150. There is poor correlation between the L index (corresponds to turbidity) and triglycerides concentration.

Lipemic specimens may cause > Abs flagging.

Drugs: No interference was found at the rapeutic concentrations using common drug panels. 8,9

Cyanokit (Hydroxocobalamin) may cause interference with results.

Physiological plasma concentrations of Sulfasalazine and Sulfapyridine may lead to false results.

In very rare cases, gammopathy, in particular type IgM (Waldenström's macroglobulinemia), may cause unreliable results.¹⁰

For diagnostic purposes, the results should always be assessed in conjunction with the patient's medical history, clinical examination and other findings.

ACTION REQUIRED

Special Wash Programming: The use of special wash steps is mandatory when certain test combinations are run together on **cobas c** systems. All special wash programming necessary for avoiding carry-over is available via the **cobas** link. The latest version of the carry-over evasion list can be found with the NaOHD/SMS/SCCS Method Sheet for information. For further instructions refer to the operator's manual.

Limits and ranges

Measuring range

5-700 U/L (0.08-11.7 µkat/L)

Determine samples having higher activities via the rerun function. Dilution of samples via the rerun function is a 1:10 dilution. Results from samples diluted using the rerun function are automatically multiplied by a factor of 10.

Lower limits of measurement

Limit of Blank, Limit of Detection and Limit of Quantitation

Limit of Blank = $5 \text{ U/L } (0.08 \, \mu \text{kat/L})$ Limit of Detection = $5 \text{ U/L } (0.08 \, \mu \text{kat/L})$ Limit of Quantitation = $10 \text{ U/L } (0.17 \, \mu \text{kat/L})$

The Limit of Blank, Limit of Detection and Limit of Quantitation were determined in accordance with the CLSI (Clinical and Laboratory Standards Institute) EP17-A2 requirements.

The Limit of Blank is the 95^{th} percentile value from $n \ge 60$ measurements of analyte-free samples over several independent series. The Limit of Blank corresponds to the activity below which analyte-free samples are found with a probability of 95%.

The Limit of Detection is determined based on the Limit of Blank and the standard deviation of low activity samples.

The Limit of Detection corresponds to the lowest analyte activity which can be detected (value above the Limit of Blank with a probability of 95 %).

The Limit of Quantitation is the lowest analyte activity that can be reproducibly measured with a total error of 20 %. It has been determined using low activity aspartate aminotransferase samples.

Expected values

U/L*

Acc. to IFCC/Standard Method 94 with pyridoxal phosphate activation measured at 37 $^{\circ}\text{C}.^{11}$

Males 10-50 U/L Females 10-35 U/L

Consensus values with pyridoxal phosphate activation: 12

Males up to 50 U/L Females up to 35 U/L

*calculated by unit conversion factor

µkat/L

Acc. to IFCC/Standard Method 94 with pyridoxal phosphate activation measured at 37 $^{\circ}\text{C}:^{11}$

Males 0.17-0.85 μkat/L Females 0.17-0.60 μkat/L



Aspartate Aminotransferase acc. to IFCC with pyridoxal phosphate activation

Consensus values with pyridoxal phosphate activation: 12

Males up to 0.85 μkat/L Females up to 0.60 μkat/L

Each laboratory should investigate the transferability of the expected values to its own patient population and if necessary determine its own reference ranges.

Specific performance data

Representative performance data on the analyzers are given below. These data represent the performance of the analytical procedure itself.

Results obtained in individual laboratories may differ due to heterogenous sample materials, aging of analyzer components and mixture of reagents running on the analyzer.

Precision

Precision was determined using human samples and controls in accordance with the CLSI (Clinical and Laboratory Standards Institute) EP05-A3 requirements with repeatability (n = 84) and intermediate precision (2 aliquots per run, 2 runs per day, 21 days). Results for repeatability and intermediate precision were obtained on the ${\bf cobas}$ ${\bf c}$ 503 analyzer.

Repeatability	Mean U/L	SD U/L	CV %
PCCC1 ^{a)}	45.7	0.651	1.4
PCCC2b)	136	0.715	0.5
Human serum 1	12.7	0.573	4.5
Human serum 2	23.9	0.809	3.4
Human serum 3	50.4	0.701	1.4
Human serum 4	344	0.932	0.3
Human serum 5	677	2.06	0.3
Intermediate precision	Mean U/L	SD U/L	CV %
Intermediate precision PCCC1 ^{a)}		~-	
•	U/L	U/L	%
PCCC1 ^{a)}	<i>U/L</i> 45.7	<i>U/L</i> 0.820	% 1.8
PCCC1 ^{a)} PCCC2 ^{b)}	<i>U/L</i> 45.7 136	<i>U/L</i> 0.820 1.47	% 1.8 1.1
PCCC1 ^{a)} PCCC2 ^{b)} Human serum 1	U/L 45.7 136 13.3	U/L 0.820 1.47 0.680	% 1.8 1.1 5.1
PCCC1 ^{a)} PCCC2 ^{b)} Human serum 1 Human serum 2	U/L 45.7 136 13.3 23.7	0.820 1.47 0.680 0.878	% 1.8 1.1 5.1 3.7

a) PreciControl ClinChem Multi 1

The data obtained on **cobas c** 503 analyzer(s) are representative for **cobas c** 303 analyzer(s).

Method comparison

AST values for human serum and plasma samples obtained on a **cobas c** 503 analyzer (y) were compared with those determined using the corresponding reagent on a **cobas c** 501 analyzer (x).

Sample size (n) = 63

 $\begin{array}{ll} Passing/Bablok^{13} & Linear\ regression \\ y = 0.979x + 0.913\ U/L & y = 0.970x + 2.53\ U/L \end{array}$

 $\tau = 0.990$ r = 1.000

The sample activities were between 5.00 and 700 U/L.

AST values for human serum and plasma samples obtained on a **cobas c** 303 analyzer (y) were compared with those determined using the corresponding reagent on a **cobas c** 501 analyzer (x).

Sample size (n) = 83

Passing/Bablok¹³ Linear regression y = 0.964x + 3.53 U/L y = 0.959x + 4.50 U/L

T = 0.989 r = 1.000

The sample activities were between 18.6 and 691 U/L.

References

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A point (period/stop) is always used in this Method Sheet as the decimal separator to mark the border between the integral and the fractional parts of a decimal numeral. Separators for thousands are not used.

Any serious incident that has occurred in relation to the device shall be reported to the manufacturer and the competent authority of the Member State in which the user and/or the patient is established.

Symbols

Roche Diagnostics uses the following symbols and signs in addition to those listed in the ISO 15223-1 standard (for USA: see dialog.roche.com for definition of symbols used):



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Additions, deletions or changes are indicated by a change bar in the margin.

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b) PreciControl ClinChem Multi 2





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