

REF		Σ	SYSTEM
07027192190	07007100500	100	cobas e 402
	07027192500	100	cobas e 801

English

System information

Short name	ACN (application code number)		
DHEAS	10068		

Intended use

Immunoassay for the in vitro quantitative determination of dehydroepiandrosterone sulfate (DHEA-S) in human serum and plasma.

The **e**lectro**c**hemiluminescence **i**mmuno**a**ssay "ECLIA" is intended for use on **cobas e** immunoassay analyzers.

Summarv

Dehydroepiandrosterone sulfate (DHEA-S) measurements, performed with this assay, in human serum and plasma are used as an aid in diagnosis and differential diagnosis of androgens related endocrine function such as hyperandrogenism, adrenal tumors and premature adrenarche.

DHEA-S is a steroid hormone synthetized in the zona reticularis of the adrenal glands in response to adrenocorticotropic hormone (ACTH). Like other steroids, DHEA-S is synthesized from cholesterol. DHEA-S is hormonally inert, but it can be converted to other more potent androgens or estrogens. Therefore DHEA-S can be considered a prohormone.

During fetal development DHEA-S is produced in the adrenal gland and the level declines rapidly during the first year of life. The production of DHEA-S resumes again during adrenarche, increases during puberty and reaches the peak values between 20 and 30 years of age. Thereafter DHEA-S levels steadily decline. In males the adrenal glands account for only a small amount of the total androgen production, while in females of reproductive age the adrenal contribution to androgen production is more pronounced.¹

Measurement of DHEA-S may be useful in the diagnostic work-up of female subjects presenting with signs and symptoms of hyperandrogenism. Hyperandrogenism is usually caused by excessive androgen production by the ovaries, the adrenal glands, or both.²

The most common disease associated with hyperandrogenism is polycystic ovary syndrome (PCOS).³ Other diseases where the measurement of DHEA-S may be useful are nonclassic congenital adrenal hyperplasia⁴, androgen-secreting adrenal tumors⁵, Cushing syndrome and hyperprolactinemia.^{1,2}

A high DHEA-S level may indicate an adrenal factor in androgen production⁶, and if substantially elevated, the presence of an adrenocortical neoplasm.⁷ DHEA-S measurements are also useful in the determination of premature adrenarche in children.⁸

The Elecsys DHEA-S assay makes use of a competition test principle using a polyclonal antibody (rabbit) specifically directed against DHEA-S. Endogenous DHEA-S in the sample competes with added DHEA-S derivative labeled with a ruthenium complex^{a)} for the binding sites on the biotinylated antibody.

a) Tris(2,2'-bipyridyl)ruthenium(II)-complex ($Ru(bpy)_3^{2+}$)

Test principle

Competition principle. Total duration of assay: 18 minutes.

- 1st incubation: By incubating the sample (9 µL) with a DHEA-S-specific biotinylated antibody, an immunocomplex is formed, the amount of which is dependent upon the analyte concentration in the sample.
- 2nd incubation: After addition of streptavidin-coated microparticles and a DHEA-S derivative labeled with a ruthenium complex, the still-vacant sites of the biotinylated antibodies become occupied, with formation of an antibody-hapten complex. The entire complex becomes bound to the solid phase via interaction of biotin and streptavidin.
- The reaction mixture is aspirated into the measuring cell where the microparticles are magnetically captured onto the surface of the electrode. Unbound substances are then removed with ProCell II M. Application of a voltage to the electrode then induces chemiluminescent emission which is measured by a photomultiplier.

 Results are determined via a calibration curve which is instrumentspecifically generated by 2-point calibration and a master curve provided via the cobas link.

Reagents - working solutions

The **cobas e** pack is labeled as DHEAS.

- M Streptavidin-coated microparticles, 1 bottle, 6.1 mL: Streptavidin-coated microparticles 0.72 mg/mL; preservative.
- R1 Anti-DHEA-S-Ab~biotin, 1 bottle, 9.9 mL: Biotinylated polyclonal anti-DHEA-S antibody (rabbit) 450 ng/mL; phosphate buffer 100 mmol/L, pH 6.8; preservative.
- R2 DHEA-S~Ru(bpy)²⁺₃, 1 bottle, 9.9 mL: DHEA-S derivative (synthetic) labeled with ruthenium complex 0.32 ng/mL; phosphate buffer 100 mmol/L, pH 6.8; preservative.

Precautions and warnings

For in vitro diagnostic use for health care professionals. Exercise the normal precautions required for handling all laboratory reagents.

Infectious or microbial waste:

Warning: handle waste as potentially biohazardous material. Dispose of waste according to accepted laboratory instructions and procedures.

Environmental hazards:

Apply all relevant local disposal regulations to determine the safe disposal. Safety data sheet available for professional user on request.

This kit contains components classified as follows in accordance with the Regulation (EC) No. 1272/2008:



Warning

H317 May cause an allergic skin reaction.

Prevention:

P261 Avoid breathing mist or vapours.

P272 Contaminated work clothing should not be allowed out of

the workplace.

P280 Wear protective gloves.

Response:

P333 + P313 If skin irritation or rash occurs: Get medical

advice/attention.

P362 + P364 Take off contaminated clothing and wash it before reuse.

Disposal:

P501 Dispose of contents/container to an approved waste

disposal plant.

Product safety labeling follows EU GHS guidance.

Contact phone: all countries: +49-621-7590

Avoid foam formation in all reagents and sample types (specimens, calibrators and controls).

Reagent handling

The reagents in the kit have been assembled into a ready-for-use unit that cannot be separated.

All information required for correct operation is available via the cobas link.



Storage and stability

Store at 2-8 °C.

Do not freeze

Store the **cobas e** pack **upright** in order to ensure complete availability of the microparticles during automatic mixing prior to use.

Stability:	
unopened at 2-8 °C	up to the stated expiration date
on the analyzers	16 weeks

Specimen collection and preparation

Only the specimens listed below were tested and found acceptable. Serum collected using standard sampling tubes or tubes containing separating gel.

Li-heparin, K₂-EDTA and K₃-EDTA plasma.

Plasma tubes containing separating gel can be used.

Criterion: Slope 0.9-1.1 + coefficient of correlation \geq 0.95.

Stable for 5 days at 20-25 °C, 14 days at 2-8 °C, 12 months at -20 °C (\pm 5 °C). Freeze only once.

The sample types listed were tested with a selection of sample collection tubes that were commercially available at the time of testing, i.e. not all available tubes of all manufacturers were tested. Sample collection systems from various manufacturers may contain differing materials which could affect the test results in some cases. When processing samples in primary tubes (sample collection systems), follow the instructions of the tube manufacturer.

Centrifuge samples containing precipitates before performing the assay. Do not use heat-inactivated samples.

Do not use samples and controls stabilized with azide.

Ensure the samples and calibrators are at 20-25 °C prior to measurement. Due to possible evaporation effects, samples and calibrators on the analyzers should be analyzed/measured within 2 hours.

Materials provided

See "Reagents – working solutions" section for reagents.

Materials required (but not provided)

- REF 03000095122, DHEA-S CalSet, for 4 x 1.0 mL
- REF 11731416190, PreciControl Universal, for 4 x 3.0 mL
- General laboratory equipment
- cobas e analyzer

Additional materials for cobas e 402 and cobas e 801 analyzers:

- REF 06908799190, ProCell II M, 2 x 2 L system solution
- REF 04880293190, CleanCell M, 2 x 2 L measuring cell cleaning solution
- REF 07485409001, Reservoir Cup, 8 cups to supply ProCell II M and CleanCell M
- REF 06908853190, PreClean II M, 2 x 2 L wash solution
- REF 05694302001, Assay Tip/Assay Cup tray, 6 magazines
 x 6 magazine stacks x 105 assay tips and 105 assay cups, 3 wasteliners
- REF 07485425001, Liquid Flow Cleaning Cup, 2 adaptor cups to supply ISE Cleaning Solution/Elecsys SysClean for Liquid Flow Cleaning Detection Unit
- REF 07485433001, PreWash Liquid Flow Cleaning Cup, 1 adaptor cup to supply ISE Cleaning Solution/Elecsys SysClean for Liquid Flow Cleaning PreWash Unit
- REF 11298500316, ISE Cleaning Solution/Elecsys SysClean, 5 x 100 mL system cleaning solution

Assav

For optimum performance of the assay follow the directions given in this document for the analyzer concerned. Refer to the appropriate operator's manual for analyzer-specific assay instructions.

Resuspension of the microparticles takes place automatically prior to use.

Place the cooled (stored at 2-8 °C) **cobas e** pack on the reagent manager. Avoid foam formation. The system automatically regulates the temperature of the reagents and the opening/closing of the **cobas e** pack.

Calibration

Traceability: This method has been standardized against gravimetrically produced master calibrators consisting of exactly defined DHEA-S concentrations in depleted human serum matrix.

The predefined master curve is adapted to the analyzer using the relevant CalSet.

Calibration frequency: Calibration must be performed once per reagent lot using fresh reagent (i.e. not more than 24 hours since the **cobas e** pack was registered on the analyzer).

Calibration interval may be extended based on acceptable verification of calibration by the laboratory.

Renewed calibration is recommended as follows:

- after 12 weeks when using the same reagent lot
- after 28 days when using the same cobas e pack on the analyzer
- as required: e.g. quality control findings outside the defined limits

Quality control

Use PreciControl Universal or other suitable controls for routine quality control procedures.

Controls for the various concentration ranges should be run individually at least once every 24 hours when the test is in use, once per **cobas e** pack, and following each calibration.

The control intervals and limits should be adapted to each laboratory's individual requirements. Values obtained should fall within the defined limits. Each laboratory should establish corrective measures to be taken if values fall outside the defined limits.

If necessary, repeat the measurement of the samples concerned. Follow the applicable government regulations and local guidelines for quality control.

Calculation

The analyzer automatically calculates the analyte concentration of each sample (either in µmol/L, µg/dL or µg/mL).

Conversion factors: $\mu mol/L \ x \ 36.846 = \mu g/dL$ $\mu g/dL \ x \ 0.02714 = \mu mol/L$ $\mu g/dL \ x \ 0.01 = \mu g/mL$

Limitations - interference

The effect of the following endogenous substances and pharmaceutical compounds on assay performance was tested. Interferences were tested up to the listed concentrations and no impact on results was observed.

Endogenous substances

Compound	Concentration tested	
Bilirubin	≤ 222 µmol/L or ≤ 13 mg/dL	
Hemoglobin	≤ 0.35 mmol/L or ≤ 0.56 g/dL	
Intralipid	≤ 2000 mg/dL	
Biotin	≤ 287 nmol/L or ≤ 70 ng/mL	
Rheumatoid factors ≤ 80 IU/mL		

Criterion: For concentrations of 0.2-50 μ g/dL the deviation is \leq \pm 5 μ g/dL. For concentrations > 50 μ g/dL the deviation is \leq \pm 10 %.

Samples should not be taken from patients receiving therapy with high biotin doses (i.e. > 5~mg/day) until at least 8 hours following the last biotin administration.

Pharmaceutical substances

In vitro tests were performed on 16 commonly used pharmaceuticals. No interference with the assay was found.

In rare cases, interference due to extremely high titers of antibodies to analyte-specific antibodies, streptavidin or ruthenium can occur. These effects are minimized by suitable test design.



For diagnostic purposes, the results should always be assessed in conjunction with the patient's medical history, clinical examination and other findings.

Limits and ranges Measuring range

 $0.005\text{-}27.1~\mu\text{mol/L}$ or $0.2\text{-}1000~\mu\text{g/dL}$ (defined by the Limit of Detection and the maximum of the master curve). Values below the Limit of Detection are reported as $<0.005~\mu\text{mol/L}$ or $<0.2~\mu\text{g/dL}$. Values above the measuring range are reported as $>27.1~\mu\text{mol/L}$ or $>1000~\mu\text{g/dL}$ (or up to 135.7 $\mu\text{mol/L}$ or 5000 $\mu\text{g/dL}$ for 5-fold diluted samples).

Lower limits of measurement

Limit of Blank, Limit of Detection and Limit of Quantitation

Limit of Blank = $0.003 \mu \text{mol/L} (0.1 \mu \text{g/dL})$

Limit of Detection = $0.005 \mu \text{mol/L}$ ($0.2 \mu \text{g/dL}$)

Limit of Quantitation = 0.081 µmol/L (3 µg/dL)

The Limit of Blank, Limit of Detection and Limit of Quantitation were determined in accordance with the CLSI (Clinical and Laboratory Standards Institute) EP17-A2 requirements.

The Limit of Blank is the 95th percentile value from $n \ge 60$ measurements of analyte-free samples over several independent series. The Limit of Blank corresponds to the concentration below which analyte-free samples are found with a probability of 95 %.

The Limit of Detection is determined based on the Limit of Blank and the standard deviation of low concentration samples. The Limit of Detection corresponds to the lowest analyte concentration which can be detected (value above the Limit of Blank with a probability of 95 %).

The Limit of Quantitation is the lowest analyte concentration that can be reproducibly measured with an intermediate precision CV of \leq 20 %.

Dilution

Samples with DHEA-S concentrations above the measuring range can be diluted using human samples with a low analyte concentration. The recommended dilution is 1:5. The concentration of the diluted sample must be > 1.22 μ mol/L (> 45 μ g/dL).

If the endogenous DHEA-S concentration is negligible, multiply the result by the dilution factor or calculate using the following equation:

C = c + 4 (c - D)

C = true DHEA-S concentration of the sample

c = measured DHEA-S concentration

D = DHEA-S concentration in the diluent (human sample)

Expected values

Extended studies with the Elecsys DHEA-S assay conducted in two clinical centers in Germany covering a total of 519 samples from female individuals, a total of 489 samples from male individuals and a total of 269 samples from children gave the following values for the age groups listed below (study protocols No.: C00P032 and C01P005 - status 05/01 to 11/01):

Age (years)	N	50 th percentile		5-95 th percentile					
		µmol/L	μg/dL	μmol/L	μg/dL				
Females:	Females:								
10-14	73	3.34	123	0.92-7.60	33.9-280				
15-19	55	4.26	157	1.77-9.99	65.1-368				
20-24	36	6.46	238	4.02-11.0	148-407				
25-34	64	4.96	183	2.68-9.23	98.8-340				
35-44*	85	4.38	161	1.65-9.15	60.9-337				
45-54*	89	3.28	121	0.96-6.95	35.4-256				
55-64	59	2.08	76.7	0.51-5.56	18.9-205				
65-74	29	1.75	64.4	0.26-6.68	9.40-246				
≥ 75	29	1.65	60.9	0.33-4.18	12.0-154				
Males:									
10-14	74	2.74	101	0.66-6.70	24.4-247				

Age (years)	N	50 th percentile		5-95 th percentile		
		μmol/L	μg/dL	μmol/L	μg/dL	
15-19	67	7.57	279	1.91-13.4	70.2-492	
20-24	28	9.58	353	5.73-13.4	211-492	
25-34	60	7.68	283	4.34-12.2	160-449	
35-44	70	6.00	221	2.41-11.6	88.9-427	
45-54	45	5.94	219	1.20-8.98	44.3-331	
55-64	69	3.75	138	1.40-8.01	51.7-295	
65-74	55	2.45	90.2	0.91-6.76	33.6-249	
≥ 75	21	1.53	56.2	0.44-3.34	16.2-123	
Children:						
< 1 week	37	7.60	280	2.93-16.5	108-607	
1-4 weeks	25	3.91	144	0.86-11.7	31.6-431	
1-12 months	69	0.59	21.6	0.09-3.35	3.4-124	
1-4 years	59	0.14	5.0	0.01-0.53	0.47-19.4	
5-9 years	79	0.63	23.1	0.08-2.31	2.8-85.2	

^{*} Effects of the menopause on the results obtained for the women of the corresponding age groups were tested and found to be negligible.

DHEA-S values of newborns are strongly influenced by maternal hormonal exchange via placenta.

Each laboratory should investigate the transferability of the expected values to its own patient population and if necessary determine its own reference ranges.

Specific performance data

Representative performance data on the analyzers are given below. Results obtained in individual laboratories may differ.

Precision

Precision was determined using Elecsys reagents, pooled human sera and controls in a protocol (EP05-A3) of the CLSI (Clinical and Laboratory Standards Institute): 2 runs per day in duplicate each for 21 days (n = 84). The following results were obtained:

cobas e 402 and cobas e 801 analyzers								
			Rep	eatabili	ty	Intermediate precision		
Sample	Mea	an	SD		CV	SD		CV
	μmol/L	μg/dL	μmol/L	μg/dL	%	μmol/L	μg/dL	%
HS ^{b)} 1	0.012	0.436	0.027	0.160	36.6	0.005	0.174	39.9
HS 2	0.131	4.82	0.015	0.539	11.2	0.018	0.667	13.8
HS 3	8.47	312	0.315	11.6	3.7	0.413	15.2	4.9
HS 4	19.0	699	0.662	24.4	3.5	0.801	29.5	4.2
HS 5	25.5	939	0.912	33.6	3.6	1.29	47.7	5.1
PC ^{c)} Univer- sal1	5.56	205	0.190	7.01	3.4	0.225	8.29	4.0
PC Univer- sal2	15.4	492	0.426	15.7	3.2	0.619	22.8	4.6

b) HS = human serumc) PC = PreciControl

Method comparison

A comparison of the Elecsys DHEA-S assay, REF 07027192190 (**cobas e** 801 analyzer; y) with the Elecsys DHEA-S assay, REF 03000087122 (**cobas e** 601 analyzer; x) gave the following correlations (μg/dL):

Number of samples measured: 148



 $\begin{array}{ll} \mbox{Passing/Bablok}^{9} & \mbox{Linear regression} \\ \mbox{y} = 0.986 \mbox{x} - 0.715 & \mbox{y} = 0.998 \mbox{x} - 3.26 \\ \mbox{\tau} = 0.971 & \mbox{r} = 0.997 \end{array}$

The sample concentrations were between 0.252 and 980 µg/dL. A comparison of the Elecsys DHEA-S assay, REF 07027192190 (cobas e 402 analyzer; y) with the Elecsys DHEA-S assay, REF 07027192190 (cobas e 801 analyzer; x) gave the following

correlations (µg/dL): Number of samples measured: 202

 Passing/Bablok9
 Linear regression

 y = 1.016x - 1.37 y = 1.006x + 1.90

 t = 0.980 t = 0.998

The sample concentrations were between 2.19 and 944 µg/dL

Analytical specificity

For the Elecsys DHEA-S assay, the following cross-reactivities were found:

Substance	Cross- reactivity %	Additive concentration µg/dL
Androstenedione	10.8	1000
DHEA	8.90	1000
Androsterone	2.10	2000
Testosterone	2.55	2000
Aldosterone	0.320	5000
Androsterone-sulfate	1.10	5000
DHEA-glucuronide	2.08	5000
Estradiol	n. d. ^{d)}	5000
Estriol	n. d.	5000
Estrone	0.740	5000
Estrone-3-sulfate	0.500	5000
Progesterone	1.32	5000
5-α-Dihydrotestosterone	1.12	5000
19-Hydroxyandrostendione	1.66	5000
Cortisol	0.060	10000

d) n. d. = not detectable

References

- Bertholf RLB, Cooper M, Winter WE. Adrenal Cortex. Rifai N, Chiu RWK, Young I, Burnham CAD, Wittwer CT, editors. Tietz Textbook of Laboratory Medicine, Saunders Elsevier, Philadelphia, 7th edition, 2023, ch. 56, p. 805-805.e37.
- 2 Goodman NF, Bledsoe MB, Cobin RH, et al. American Association of Clinical Endocrinologists Hyperandrogenic Disorders Task Force. American Association of Clinical Endocrinologists medical guidelines for the clinical practice for the diagnosis and treatment of hyperandrogenic disorders. Endocr Pract. 2001 Mar-Apr;7(2):120-34.
- Teede HJ, Tay CT, Laven J, et al. International PCOS Network. Recommendations from the 2023 International Evidence-based Guideline for the Assessment and Management of Polycystic Ovary Syndrome†. Hum Reprod. 2023 Sep 5;38(9):1655-1679.
- 4 Speiser PW, Arlt W, Auchus RJ, et al. Congenital Adrenal Hyperplasia Due to Steroid 21-Hydroxylase Deficiency: An Endocrine Society Clinical Practice Guideline. J Clin Endocrinol Metab. 2018 Nov 1;103(11):4043-4088.
- Fassnacht M, Dekkers OM, Else T, et al. European Society of Endocrinology Clinical Practice Guidelines on the management of adrenocortical carcinoma in adults, in collaboration with the European Network for the Study of Adrenal Tumors. Eur J Endocrinol. 2018 Oct 1;179(4):G1-G46.

- 6 Goodarzi MO, Carmina E, Azziz R. DHEA, DHEAS and PCOS. J Steroid Biochem Mol Biol. 2015 Jan;145:213-25.
- 7 Pugeat M, Déchaud H, Raverot V, et al. Recommendations for investigation of hyperandrogenism. Ann Endocrinol (Paris). 2010 Feb;71(1):2-7.
- 8 Rosenfield RL. Normal and Premature Adrenarche. Endocr Rev. 2021 Nov 16;42(6):783-814.
- 9 Bablok W, Passing H, Bender R, et al. A general regression procedure for method transformation. Application of linear regression procedures for method comparison studies in clinical chemistry, Part III. J Clin Chem Clin Biochem 1988 Nov;26(11):783-790.

For further information, please refer to the appropriate user guide or operator's manual for the analyzer concerned, the respective application sheets and the Method Sheets of all necessary components (if available in your country).

A point (period/stop) is always used in this Method Sheet as the decimal separator to mark the border between the integral and the fractional parts of a decimal numeral. Separators for thousands are not used.

Any serious incident that has occurred in relation to the device shall be reported to the manufacturer and the competent authority of the Member State in which the user and/or the patient is established.

Symbols

Roche Diagnostics uses the following symbols and signs in addition to those listed in the ISO 15223-1 standard (for USA: see navifyportal.roche.com for definition of symbols used):

CONTENT Contents of kit

SYSTEM Analyzers/Instruments on which reagents can be used

REAGENT Reagent

CALIBRATOR Calibrator

Volume for reconstitution

GTIN Global Trade Item Number

Rx only For USA: Caution: Federal law restricts this device to

sale by or on the order of a physician.

COBAS, NAVIFY, ELECSYS and PRECICONTROL are trademarks of Roche. INTRALIPID is a trademark of Fresenius Kabi AB.

All other product names and trademarks are the property of their respective owners Additions, deletions or changes are indicated by a change bar in the margin. © 2023, Roche Diagnostics

(€ 0123

Roche Diagnostics GmbH, Sandhofer Strasse 116, D-68305 Mannheim

+800 5505 6606

