

#### **Rheumatoid Factors II**

#### Order information



REF	(i)	CONTENT		Analyzer(s) on which <b>cobas c</b> pack(s) can be used
08058628190	08058628500	Rheumatoid Factors II (400 tests)	System-ID 2104 001	cobas c 303, cobas c 503, cobas c 703

Materials required (but not provided):

12172828322	Preciset RF (5 × 1 mL)	Codes 20725-20729	
	RF Control Set		
03005496122	Level I (2 x 1 mL)	Code 20215	
	Level II (2 x 1 mL)	Code 20216	
08063494190	Diluent NaCl 9 % (123 mL)	System-ID 2906 001	

#### **English**

# System information RF-II: ACN 21040

#### Intended use

In vitro test for the quantitative determination of Rheumatoid Factors (RF-II) in human serum and plasma on **cobas c** systems. Measurements may be used as an aid in the diagnosis of rheumatoid arthritis.

#### Summary

Measurements of rheumatoid factors with this assay in human serum and plasma may be used as an aid in the diagnosis of rheumatoid arthritis.

Rheumatoid factors are a heterogeneous group of autoantibodies directed against the antigenic determinants on the Fc-region of IgG molecules.¹ They are important in the diagnosis of rheumatoid arthritis, but can also be found in other inflammatory rheumatic diseases and in various non-rheumatic diseases. They are also found in clinically healthy persons, however with low or moderate levels. Despite these restrictions, the detection of rheumatoid factors is a diagnostic criterion in several clinical guidelines for classifying rheumatoid arthritis.¹2,3,4,5

The autoantibodies occur in all the immunoglobulin classes, although the usual analytical methods are limited to the detection of rheumatoid factors of the IgM type. The classic procedure for the quantitation of rheumatoid factors is by agglutination with IgG-sensitized sheep erythrocytes or latex particles. Particular problems of these semiquantitative methods are the poor between-laboratory precision and reproducibility, together with standardization difficulties. For these reasons, new assay methods such as nephelometry, turbidimetry, enzyme-immunoassays and radioimmunoassays have been developed. 6.7.8 This assay is based on the immunological agglutination principle with enhancement of the reaction by latex.

# Test principle<sup>1,9,10</sup>

Immunoturbidimetric assay

Latex-bound heat-inactivated IgG (antigen) reacts with the RF-antibodies in the sample to form antigen/antibody complexes which, following agglutination, are measured turbidimetrically.

### Reagents - working solutions

R1 Glycine buffer: 170 mmol/L, pH 8.0; polyethylene glycol: 0.05 %; bovine serum albumin; stabilizer; preservative

R3 Latex particles coated with human IgG; glycine buffer: 170 mmol/L, pH 7.3; stabilizer; preservative

R1 is in position B and R3 is in position C.

#### Precautions and warnings

For in vitro diagnostic use for health care professionals. Exercise the normal precautions required for handling all laboratory reagents.

Infectious or microbial waste:

Warning: handle waste as potentially biohazardous material. Dispose of waste according to accepted laboratory instructions and procedures.

Environmental hazards:

Apply all relevant local disposal regulations to determine the safe disposal. Safety data sheet available for professional user on request.

This kit contains components classified as follows in accordance with the Regulation (EC) No. 1272/2008:



#### Warning

H317 May cause an allergic skin reaction.

Prevention:

P261 Avoid breathing mist or vapours.

P272 Contaminated work clothing should not be allowed out of

the workplace.

P280 Wear protective gloves.

Response:

P333 + P313 If skin irritation or rash occurs: Get medical

advice/attention.

P362 + P364 Take off contaminated clothing and wash it before reuse.

Disposal:

P501 Dispose of contents/container to an approved waste

disposal plant.

Product safety labeling follows EU GHS guidance.

Contact phone: all countries: +49-621-7590

All human material should be considered potentially infectious. All products derived from human blood are prepared exclusively from the blood of donors tested individually and shown to be free from HBsAg and antibodies to HCV and HIV. The testing methods use assays that have been approved or cleared by the FDA or that are in compliance with the legal rules of the European Union (IVDR 2017/746/EU, IVDD 98/79/EC, Annex II, List A). However, as no testing method can rule out the potential risk of infection with absolute certainty, the material should be handled with the same level of care as a patient specimen. In the event of exposure, the directives of the responsible health authorities should be followed. 11.12

### Reagent handling

Ready for use

Carefully invert reagent container several times prior to use to ensure that the reagent components are mixed.

#### Storage and stability

Shelf life at 2-8 °C: See expiration date on **cobas c** pack label.

On-board in use and refrigerated on the

8 weeks

analyzer:



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#### Specimen collection and preparation

For specimen collection and preparation only use suitable tubes or collection containers.

Only the specimens listed below were tested and found acceptable. Serum

Plasma: Li-heparin and K<sub>2</sub>-EDTA plasma

The sample types listed were tested with a selection of sample collection tubes that were commercially available at the time of testing, i.e. not all available tubes of all manufacturers were tested. Sample collection systems from various manufacturers may contain differing materials which could affect the test results in some cases. When processing samples in primary tubes (sample collection systems), follow the instructions of the tube manufacturer.

Centrifuge samples containing precipitates before performing the assay. See the limitations and interferences section for details about possible sample interferences.

Stability:13 1 day at 20-25 °C

8 days at 4-8 °C

3 months at -20 °C (±5 °C)

Freeze only once.

#### Materials provided

See "Reagents - working solutions" section for reagents.

#### Materials required (but not provided)

See "Order information" section

General laboratory equipment

#### Assay

For optimum performance of the assay follow the directions given in this document for the analyzer concerned. Refer to the appropriate operator's manual for analyzer-specific assay instructions.

The performance of applications not validated by Roche is not warranted and must be defined by the user.

# Application for serum and plasma

#### **Test definition**

Reporting time	10 min
Wavelength (sub/main)	800/570 nm
Reagent pipetting	

R1 63 µL

R3 21 µL

Sample volumes	Sample	Sam	ple dilution
		Sample	Diluent (NaCl
Normal	2.1 μL	-	_
Decreased	2.1 μL	20 μL	80 μL
Increased	2.1 µL	_	_

For further information about the assay test definitions refer to the application parameters screen of the corresponding analyzer and assay.

#### Calibration

S1: H<sub>2</sub>O Calibrators

S2-6: Preciset RF

Calibration mode Non-linear Calibration frequency Full calibration

> - every 180 days during shelf life - after reagent lot change

- as required following quality control

Diluent (H2O)

procedures

Calibration interval may be extended based on acceptable verification of calibration by the laboratory.

Traceability: 14 This method has been standardized using the WHO Standard 64/2.

#### **Quality control**

For quality control, use control materials as listed in the "Order information" section. In addition, other suitable control material can be used

The control intervals and limits should be adapted to each laboratory's individual requirements. It is recommended to perform quality control always after lot calibration and subsequently at least every 8 weeks. Values obtained should fall within the defined limits. Each laboratory should establish corrective measures to be taken if values fall outside the defined

Follow the applicable government regulations and local guidelines for quality control.

#### Calculation

cobas c systems automatically calculate the analyte concentration of each sample in the unit IU/mL

#### **Limitations - interference**

Criterion: Recovery within ± 1.4 IU/mL of initial values of samples ≤ 14 IU/mL and within ± 10 % for samples > 14 IU/mL.

Icterus: 15 No significant interference up to an I index of 40 for conjugated and 60 for unconjugated bilirubin (approximate conjugated bilirubin concentration: 624 µmol/L or 40 mg/dL and approximate unconjugated bilirubin concentration: 1026 µmol/L or 60 mg/dL).

Hemolysis:<sup>15</sup> No significant interference up to an H index of 300 (approximate hemoglobin concentration: 186 µmol/L or 300 mg/dL).

Lipemia (Intralipid):<sup>15</sup> No significant interference up to an L index of 2000. There is poor correlation between the L index (corresponds to turbidity) and triglycerides concentration.

Drugs: No interference was found at the rapeutic concentrations using common drug panels. 16,17

High-dose hook effect: Using the prozone check automatically performed by the analyzer, no false result without a flag was observed up to an RF concentration of 6000 IU/mL.

In very rare cases, gammopathy, in particular type IgM (Waldenström's macroglobulinemia), may cause unreliable results.  $^{\rm 18}$ 

There is the possibility that other substances and/or factors may interfere with the test and cause unreliable results.

For diagnostic purposes, the results should always be assessed in conjunction with the patient's medical history, clinical examination and other findings.

# **ACTION REQUIRED**

Special Wash Programming: The use of special wash steps is mandatory when certain test combinations are run together on cobas c systems. All special wash programming necessary for avoiding carry-over is available via the **cobas** link. The latest version of the carry-over evasion list can be found with the NaOHD/SMS/SCCS Method Sheet. For further instructions, refer to the operator's manual.

# Limits and ranges

# Measuring range

10-130 IU/mL

Determine samples having higher concentrations via the rerun function. Dilution of samples via the rerun function is a 1:5 dilution. Results from samples diluted using the rerun function are automatically multiplied by a factor of 5

# Lower limits of measurement

Limit of Blank, Limit of Detection and Limit of Quantitation

= 10 IU/mL Limit of Blank = 10 IU/mL Limit of Detection = 10 IU/mL Limit of Quantitation

The Limit of Blank, Limit of Detection and Limit of Quantitation were determined in accordance with the CLSI (Clinical and Laboratory Standards Institute) EP17-A2 requirements.



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The Limit of Blank is the  $95^{th}$  percentile value from  $n \ge 60$  measurements of analyte-free samples over several independent series. The Limit of Blank corresponds to the concentration below which analyte-free samples are found with a probability of 95%.

The Limit of Detection is determined based on the Limit of Blank and the standard deviation of low concentration samples.

The Limit of Detection corresponds to the lowest analyte concentration which can be detected (value above the Limit of Blank with a probability of 95%).

The Limit of Quantitation is the lowest analyte concentration that can be reproducibly measured with a total error of 20 %. It has been determined using low concentration RF samples.

#### **Expected values**

< 14 IU/mL

This value is based on serum samples from 541 test subjects.

Each laboratory should investigate the transferability of the expected values to its own patient population and if necessary determine its own reference ranges.

#### Specific performance data

Representative performance data on the analyzers are given below. These data represent the performance of the analytical procedure itself.

Results obtained in individual laboratories may differ due to heterogenous sample materials, aging of analyzer components and mixture of reagents running on the analyzer.

#### Precision

Precision was determined using human samples and controls in accordance with the CLSI (Clinical and Laboratory Standards Institute) EP05-A3 requirements with repeatability (n = 84) and intermediate precision (2 aliquots per run, 2 runs per day, 21 days). Results for repeatability and intermediate precision were obtained on the **cobas c** 503 analyzer.

Repeatability	Mean IU/mL	SD IU/mL	CV %
RF Control Level 1	21.8	0.509	2.3
RF Control Level 2	51.3	0.329	0.6
Human serum 1	15.0	0.704	4.7
Human serum 2	30.2	0.295	1.0
Human serum 3	37.9	0.250	0.7
Human serum 4	63.0	0.286	0.5
Human serum 5	109	0.707	0.6
Intermediate precision	Mean IU/mL	SD IU/mL	CV %
Intermediate precision  RF Control Level 1			
•	IU/mL	IU/mL	%
RF Control Level 1	<i>IU/mL</i> 21.8	<i>IU/mL</i> 0.550	% 2.5
RF Control Level 1 RF Control Level 2	IU/mL 21.8 51.3	IU/mL 0.550 0.408	% 2.5 0.8
RF Control Level 1 RF Control Level 2 Human serum 1	IU/mL 21.8 51.3 15.0	IU/mL 0.550 0.408 0.704	% 2.5 0.8 4.7
RF Control Level 1 RF Control Level 2 Human serum 1 Human serum 2	1U/mL 21.8 51.3 15.0 30.2	IU/mL 0.550 0.408 0.704 0.396	% 2.5 0.8 4.7 1.3

The data obtained on **cobas c** 503 analyzer(s) are representative for **cobas c** 303 analyzer(s) and **cobas c** 703 analyzer(s).

#### Method comparison

RF values for human serum and plasma samples obtained on a **cobas c** 503 analyzer (y) were compared with those determined using the corresponding reagent on a **cobas c** 501 analyzer (x).

Sample size (n) = 68

 $\begin{array}{ll} Passing/Bablok^{19} & Linear\ regression \\ y = 1.004x + 0.00648\ IU/mL & y = 0.986x + 0.494\ IU/mL \\ \tau = 0.949 & r = 0.996 \end{array}$ 

The sample concentrations were between 12.6 and 121 IU/mL.

RF values for human serum and plasma samples obtained on a **cobas c** 303 analyzer (y) were compared with those determined using the corresponding reagent on a **cobas c** 501 analyzer (x).

Sample size (n) = 65

Passing/Bablok<sup>19</sup> Linear regression y = 1.037x - 1.61 IU/mL y = 1.016x - 0.990 IU/mL t = 0.942 t = 0.995

The sample concentrations were between 11.8 and 125 IU/mL.

RF values for human serum and plasma samples obtained on a **cobas c** 703 analyzer (y) were compared with those determined using the corresponding reagent on a **cobas c** 503 analyzer (x).

Sample size (n) = 66

Passing/Bablok<sup>19</sup> Linear regression y = 1.026x - 1.70 IU/mL y = 1.027x - 1.70 IU/mL

T = 0.971 r = 0.999

The sample concentrations were between 11.4 and 130 IU/mL.

A point (period/stop) is always used in this Method Sheet as the decimal separator to mark the border between the integral and the fractional parts of a decimal numeral. Separators for thousands are not used.

Any serious incident that has occurred in relation to the device shall be reported to the manufacturer and the competent authority of the Member State in which the user and/or the patient is established.

#### References

- Moore TL, Dorner RW. Rheumatoid factors. Clin Biochem 1993 Apr;26(2):75-84.
- Ingegnoli F, Castelli R, Gualtierotti R. Rheumatoid factors: clinical applications. Dis Markers 2013;35(6):727-734.
- 3 Aletaha D, Neogi T, Silman AJ, et al. 2010 rheumatoid arthritis classification criteria: an American College of Rheumatology/European League Against Rheumatism collaborative initiative. Ann Rheum Dis 2010 Sep;69(9):1580-1588.
- 4 Combe B, Landewe R, Daien CI, et al. 2016 update of the EULAR recommendations for the management of early arthritis. Ann Rheum Dis 2017 Jun;76(6):948-959.
- National Institute for Health and Care Excellence (NICE) (2018). Rheumatoid arthritis in adults: diagnosis and management. (NICE Guideline [NG100]) [updated 2020 October; cited 2024 February 15]. Available from: https://www.nice.org.uk/guidance/ng100.
- 6 Bampton JL, Cawston TE, Kyle MV, et al. Measurement of rheumatoid factors by an enzyme-linked immunosorbent assay (ELISA) and comparison with other methods. Ann Rheum Dis 1985 Jan;44(1):13-19.
- 7 Koopman WJ, Schrohenloher RE. A sensitive radioimmunoassay for quantitation of IgM rheumatoid factor. Arthritis Rheum 1980 Mar:23(3):302-308.
- 8 Jaspers JP, Van Oers RJ, Leerkes B. Nine rheumatoid factor assays compared. J Clin Chem Clin Biochem 1988 Dec;26(12):863-871.
- 9 Waaler E. On the occurrence of a factor in human serum activating the specific agglutination of sheep blood corpuscles. Acta Pathol Microbiol Scand 1940;17:172-178.
- Roberts-Thomson PJ, McEvoy R, et al. Ann Rheum Dis 1985;44:379-383.
- 11 Occupational Safety and Health Standards: Bloodborne pathogens. (29 CFR Part 1910.1030). Fed. Register.
- 12 Directive 2000/54/EC of the European Parliament and Council of 18 September 2000 on the protection of workers from risks related to exposure to biological agents at work.
- 13 Use of Anticoagulants in Diagnostic Laboratory Investigations. WHO Publication WHO/DIL/LAB/99.1 Rev. 2: Jan 2002.
- 14 Anderson SG, Bentzon MW, Houba V, et al. International reference preparation of rheumatoid arthritis serum. Bull Wld Hlth Org 1970;42:311-318.

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- 15 Glick MR, Ryder KW, Jackson SA. Graphical Comparisons of Interferences in Clinical Chemistry Instrumentation. Clin Chem 1986;32:470-475.
- Breuer J. Report on the Symposium "Drug effects in Clinical Chemistry Methods". Eur J Clin Chem Clin Biochem 1996;34:385-386.
- 17 Sonntag O, Scholer A. Drug interference in clinical chemistry: recommendation of drugs and their concentrations to be used in drug interference studies. Ann Clin Biochem 2001;38:376-385.
- 18 Bakker AJ, Mücke M. Gammopathy interference in clinical chemistry assays: mechanisms, detection and prevention. Clin Chem Lab Med 2007;45(9):1240-1243.
- 19 Bablok W, Passing H, Bender R, et al. A general regression procedure for method transformation. Application of linear regression procedures for method comparison studies in clinical chemistry, Part III. J Clin Chem Clin Biochem 1988 Nov;26(11):783-790.

#### Symbols

Roche Diagnostics uses the following symbols and signs in addition to those listed in the ISO 15223-1 standard:



Contents of kit

Volume for reconstitution

Global Trade Item Number

Rx only

For USA: Caution: Federal law restricts this device to sale by or on the order of a

physician.

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