08429324500V3.0

Elecsys TSH

cobas®

REF

08429324190

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English

System information

For **cobas e** 411 analyzer: test number 1820 For **cobas e** 601 and **cobas e** 602 analyzers: Application Code Number 627

Intended use

Immunoassay for the in vitro quantitative determination of thyrotropin in human serum and plasma.

The electrochemiluminescence immunoassay "ECLIA" is intended for use on **cobas e** immunoassay analyzers.

Summary

Thyroid-stimulating hormone (TSH, thyrotropin) measurements, performed with this assay, in human serum and plasma are used as an aid in diagnosis of hypothalamic-pituitary-thyroid (HPT) disorders.

TSH is a glycoprotein consisting of 2 subunits. The β -subunit is critical for the heterodimer assembly and its biological activity. The α -chain is common to the follicle-stimulating hormone (FSH), luteinizing hormone (LH), and human chorionic gonadotropin (hCG).¹

TSH is formed in specific basophil cells of the anterior pituitary and is subject to a circadian secretion sequence. The hypophyseal synthesis and release of TSH is stimulated by the thyrotropin-releasing hormone (TRH). TSH binding to the TSH receptor results in the production of the thyroid hormones triiodothyronine (T3) and thyroxine (T4). T3 and T4 in turn regulate the TSH and TRH levels via a negative feedback loop. Small changes in the concentrations of free T3 and/or T4 will result in significant large changes in TSH concentrations.²

The determination of TSH serves as the initial test to assess thyroid function. In case of high levels of TSH, free thyroxine (fT4) measurements are performed to determine the degree of hypothyroidism. When low levels of TSH are measured, fT4 and free triiodothyronine (fT3) measurements are performed to determine the degree of hyperthyroidism. TSH measurements are performed to determine the degree of arealy detection or exclusion of disorders in the central regulating circuit between the hypothalamus, pituitary and thyroid. In case of suspected pituitary or hypothalamic disease, TSH levels are measured together with fT4 levels to confirm diagnosis.^{3,4,5,6}

The Elecsys TSH assay employs monoclonal antibodies specifically directed against human TSH. The antibodies labeled with ruthenium complex^{a)} consist of a chimeric construct from human and mouse-specific components. As a result, interfering effects due to HAMA (human anti-mouse antibodies) are largely eliminated.

a) Tris(2,2'-bipyridyl)ruthenium(II)-complex (Ru(bpy)₃²⁺)

Test principle

Sandwich principle. Total duration of assay: 18 minutes.

- 1st incubation: 50 µL of sample, a biotinylated monoclonal TSH-specific antibody and a monoclonal TSH-specific antibody labeled with a ruthenium complex react to form a sandwich complex.
- 2nd incubation: After addition of streptavidin-coated microparticles, the complex becomes bound to the solid phase via interaction of biotin and streptavidin.
- The reaction mixture is aspirated into the measuring cell where the microparticles are magnetically captured onto the surface of the electrode. Unbound substances are then removed with ProCell/ProCell M. Application of a voltage to the electrode then induces chemiluminescent emission which is measured by a photomultiplier.
- Results are determined via a calibration curve which is instrumentspecifically generated by 2-point calibration and a master curve provided via the reagent barcode or e-barcode.

Reagents - working solutions

The reagent rackpack is labeled as TSH.

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	со	obas e 602		
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SYSTEM

- M Streptavidin-coated microparticles (transparent cap), 1 bottle, 12 mL: Streptavidin-coated microparticles 0.72 mg/mL; preservative.
- R1 Anti-TSH-Ab~biotin (gray cap), 1 bottle, 14 mL: Biotinylated monoclonal anti-TSH antibody (mouse) 2.0 mg/L; phosphate buffer 100 mmol/L, pH 7.2; preservative.
- R2 Anti-TSH-Ab~Ru(bpy)²⁺₃ (black cap), 1 bottle, 12 mL: Monoclonal anti-TSH antibody (mouse/human) labeled with ruthenium complex 1.5 mg/L; phosphate buffer 100 mmol/L, pH 7.2; preservative.

Precautions and warnings

Σ

For in vitro diagnostic use for health care professionals. Exercise the normal precautions required for handling all laboratory reagents. Infectious or microbial waste:

Warning: handle waste as potentially biohazardous material. Dispose of waste according to accepted laboratory instructions and procedures. Environmental hazards:

Apply all relevant local disposal regulations to determine the safe disposal. Safety data sheet available for professional user on request.

This kit contains components classified as follows in accordance with the Regulation (EC) No. $1272/2008\colon$



Warning

H317 May cause an allergic skin reaction.

Prevention:

- P261 Avoid breathing mist or vapours.
- P272 Contaminated work clothing should not be allowed out of the workplace.
- P280 Wear protective gloves.

Response:

P333 + P313 If skin irritation or rash occurs: Get medical advice/attention.

P362 + P364 Take off contaminated clothing and wash it before reuse. **Disposal:**

P501 Dispose of contents/container to an approved waste disposal plant.

Product safety labeling follows EU GHS guidance.

Contact phone: all countries: +49-621-7590

Avoid foam formation in all reagents and sample types (specimens, calibrators and controls).

Reagent handling

The reagents in the kit have been assembled into a ready-for-use unit that cannot be separated.

All information required for correct operation is read in from the respective reagent barcodes.

Storage and stability

Store at 2-8 °C. Do not freeze.

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Store the Elecsys reagent kit **upright** in order to ensure complete availability of the microparticles during automatic mixing prior to use.

Stability:

unopened at 2-8 °C	up to the stated expiration date
after opening at 2-8 °C	12 weeks
on cobas e 601 and cobas e 602	6 weeks
on cobas e 411	8 weeks
	0

Specimen collection and preparation

Only the specimens listed below were tested and found acceptable. Serum collected using standard sampling tubes or tubes containing separating gel.

Li-heparin, K₂-EDTA and K₃-EDTA plasma.

Plasma tubes containing separating gel can be used.

Criterion: Slope 0.9-1.1 + coefficient of correlation \ge 0.95 and with a bias \le 10 % at medical decision points (0.27 µIU/mL and 4.2 µIU/mL).

Stable for 8 days at 20-25 $^{\circ}\text{C},$ 14 days at 2-8 $^{\circ}\text{C},$ 24 months at -20 $^{\circ}\text{C}$ (± 5 $^{\circ}\text{C}). Freeze only once.$

The sample types listed were tested with a selection of sample collection tubes that were commercially available at the time of testing, i.e. not all available tubes of all manufacturers were tested. Sample collection systems from various manufacturers may contain differing materials which could affect the test results in some cases. When processing samples in primary tubes (sample collection systems), follow the instructions of the tube manufacturer.

Centrifuge samples containing precipitates before performing the assay.

Do not use heat-inactivated samples.

Do not use samples and controls stabilized with azide.

Ensure the samples, calibrators and controls are at 20-25 $^\circ \text{C}$ prior to measurement.

Due to possible evaporation effects, samples, calibrators and controls on the analyzers should be analyzed/measured within 2 hours.

Materials provided

See "Reagents - working solutions" section for reagents.

Materials required (but not provided)

- REF 08443459190, TSH CalSet, 4 x 1.3 mL
- REF 11731416190, PreciControl Universal, for 4 x 3.0 mL
- REF 06445918190, PreciControl Thyro Sensitive, for 4 x 2.0 mL
- REF 03609987190, Diluent MultiAssay, 2 x 16 mL sample diluent
- General laboratory equipment

cobas e analyzer

Additional materials for the **cobas e** 411 analyzer:

- REF 11662988122, ProCell, 6 x 380 mL system buffer
- REF 11662970122, CleanCell, 6 x 380 mL measuring cell cleaning solution
- REF 11930346122, Elecsys SysWash, 1 x 500 mL washwater additive
- REF 11933159001, Adapter for SysClean
- REF 11706802001, AssayCup, 60 x 60 reaction cups
- REF 11706799001, AssayTip, 30 x 120 pipette tips
- REF 11800507001, Clean-Liner
- Additional materials for cobas e 601 and cobas e 602 analyzers:
- REF 04880340190, ProCell M, 2 x 2 L system buffer
- REF 04880293190, CleanCell M, 2 x 2 L measuring cell cleaning solution
- REF 03023141001, PC/CC-Cups, 12 cups to prewarm ProCell M and CleanCell M before use
- REF 03005712190, ProbeWash M, 12 x 70 mL cleaning solution for run finalization and rinsing during reagent change



- REF 12102137001, AssayTip/AssayCup, 48 magazines x 84 reaction cups or pipette tips, waste bags
- REF 03023150001, WasteLiner, waste bags
- REF 03027651001, SysClean Adapter M

Additional materials for all analyzers:

 REF 11298500316, ISE Cleaning Solution/Elecsys SysClean, 5 x 100 mL system cleaning solution

Assay

For optimum performance of the assay follow the directions given in this document for the analyzer concerned. Refer to the appropriate operator's manual for analyzer-specific assay instructions.

Resuspension of the microparticles takes place automatically prior to use. Read in the test-specific parameters via the reagent barcode. If in exceptional cases the barcode cannot be read, enter the 15-digit sequence of numbers.

Bring the cooled reagents to approximately 20 °C and place on the reagent disk (20 °C) of the analyzer. Avoid foam formation. The system automatically regulates the temperature of the reagents and the opening/closing of the bottles.

Calibration

Traceability: This method has been standardized against the 2nd IRP WHO Reference Standard 80/558.

Every Elecsys reagent set has a barcoded label containing specific information for calibration of the particular reagent lot. The predefined master curve is adapted to the analyzer using the relevant CalSet.

Calibration frequency: Calibration must be performed once per reagent lot using fresh reagent (i.e. not more than 24 hours since the reagent kit was registered on the analyzer).

Calibration interval may be extended based on acceptable verification of calibration by the laboratory.

Renewed calibration is recommended as follows:

- after 8 weeks when using the same reagent lot
- after 7 days when using the same reagent kit on the analyzer
- as required: e.g. quality control findings outside the defined limits

Quality control

For quality control, use PreciControl Universal or PreciControl Thyro Sensitive.

In addition, other suitable control material can be used.

Controls for the various concentration ranges should be run individually at least once every 24 hours when the test is in use, once per reagent kit, and following each calibration.

The control intervals and limits should be adapted to each laboratory's individual requirements. Values obtained should fall within the defined limits. Each laboratory should establish corrective measures to be taken if values fall outside the defined limits.

If necessary, repeat the measurement of the samples concerned.

Follow the applicable government regulations and local guidelines for quality control.

Calculation

The analyzer automatically calculates the analyte concentration of each sample either in μ IU/mL or mIU/L (selectable).

Limitations - interference

The effect of the following endogenous substances and pharmaceutical compounds on assay performance was tested. Interferences were tested up to the listed concentrations and no impact on results was observed.

Endogenous substances

Compound	Concentration tested
Bilirubin	\leq 701 µmol/L or \leq 41 mg/dL
Hemoglobin	≤ 0.621 mmol/L or ≤ 1000 mg/dL
Intralipid	≤ 1500 mg/dL
Biotin	≤ 4912 nmol/L or ≤ 1200 ng/mL

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Compound	Concentration tested
Rheumatoid factors	≤ 1500 IU/mL
IgG	≤ 2 g/dL
IgM	≤ 0.5 g/dL

Criterion: For concentrations \leq 0.2 µIU/mL the deviation is \leq 0.02 µIU/mL. For concentrations > 0.2 µIU/mL the deviation is \leq 10 %.

There is no high-dose hook effect at TSH concentrations up to 1000 $\mu\text{IU/mL}.$

Pharmaceutical substances

In vitro tests were performed on 17 commonly used pharmaceuticals. No interference with the assay was found.

In addition, the following special drugs were tested. No interference with the assay was found.

Special drugs

Drug	Concentration tested mg/L
lodide	0.2
Carbimazole	30
Methimazole	80
Propylthiouracil	60
Perchlorate	2000
Propranolol	240
Amiodarone	200
Prednisolone	100
Hydrocortisone	200
Fluocortolone	100
Octreotide	0.3
Levothyroxine	0.25
Liothyronine	0.015

Drug interferences are measured based on recommendations given in CLSI guidelines EP07 and EP37 and other published literature. Effects of concentrations exceeding these recommendations have not been characterized.

The presence of autoantibodies may induce high molecular weight complexes (macro-TSH) which may cause unexpectedly high values of TSH. $^{7}\,$

In rare cases, interference due to extremely high titers of antibodies to analyte-specific antibodies, streptavidin or ruthenium can occur. These effects are minimized by suitable test design.

For diagnostic purposes, the results should always be assessed in conjunction with the patient's medical history, clinical examination and other findings.

Limits and ranges

Measuring range

 $0.005-100 \mu$ IU/mL (defined by the Limit of Detection and the maximum of the master curve). Values below the Limit of Detection are reported as < 0.005μ IU/mL. Values above the measuring range are reported as > 100μ IU/mL (or up to 1000μ IU/mL for 10-fold diluted samples).

Lower limits of measurement

Limit of Blank, Limit of Detection and Limit of Quantitation

Limit of Blank = 0.0025 µIU/mL

Limit of Detection = 0.005 μ IU/mL

Limit of Quantitation = 0.005 µIU/mL

The Limit of Blank, Limit of Detection and Limit of Quantitation were determined in accordance with the CLSI (Clinical and Laboratory Standards Institute) EP17-A2 requirements.

The Limit of Blank is the 95^{in} percentile value from $n \ge 60$ measurements of analyte-free samples over several independent series. The Limit of Blank

corresponds to the concentration below which analyte-free samples are found with a probability of 95 %.

The Limit of Detection is determined based on the Limit of Blank and the standard deviation of low concentration samples. The Limit of Detection corresponds to the lowest analyte concentration which can be detected (value above the Limit of Blank with a probability of 95 %).

The Limit of Quantitation is the lowest analyte concentration that can be reproducibly measured with an intermediate precision CV of \leq 20 %.

Dilution

Samples with TSH concentrations above the measuring range can be diluted with Diluent MultiAssay. The recommended dilution is 1:10 (either automatically by the analyzers, or manually). The concentration of the diluted sample must be \geq 10 μ IU/mL.

After manual dilution, multiply the result by the dilution factor.

After dilution by the analyzers, the software automatically takes the dilution into account when calculating the sample concentration.

Expected values

0.270-4.20 µIU/mL8

These values correspond to the 2.5^{th} and 97.5^{th} percentiles of results obtained from a total of 516 healthy test subjects examined.

For detailed information about reference intervals in children, adolescents and pregnant women, refer to the brochure "Reference Intervals for Children and Adults", English: [REF] 04640292.

This booklet also contains results of a detailed study about influencing factors on thyroid parameters in a well characterized reference group of adults. Different inclusion and exclusion criteria were applied (e.g. sonographic results (thyroid volume and density) as well as criteria according to the guidelines of the National Academy of Clinical Biochemistry - NACB).

Each laboratory should investigate the transferability of the expected values to its own patient population and if necessary determine its own reference ranges.

Specific performance data

Representative performance data on the analyzers are given below. Results obtained in individual laboratories may differ.

Precision

Precision was determined using Elecsys reagents, samples and controls in a protocol (EP05-A3) of the CLSI (Clinical and Laboratory Standards Institute): 2 runs per day in duplicate each for 21 days (n = 84). The following results were obtained:

cobas e 411 analyzer					
		Repeatability		Intermediate precision	
Sample	Mean µIU/mL	SD µIU/mL	CV %	SD µIU/mL	CV %
Human serum 1	0.016	0.001	4.2	0.001	6.3
Human serum 2	0.277	0.004	1.5	0.013	4.7
Human serum 3	3.82	0.056	1.5	0.177	4.6
Human serum 4	53.8	0.807	1.5	2.18	4.0
Human serum 5	93.6	1.62	1.7	2.23	2.4
PC ^{b)} Universal 1	1.42	0.016	1.2	0.058	4.1
PC Universal 2	7.97	0.137	1.7	0.307	3.8
PC Thyro Sensitive	0.179	0.003	1.5	0.009	5.2

b) PC = PreciControl

cobas e 601 and cobas e 602 analyzers					
		Repeatability Intermediate			
Sample	Mean µIU/mL	SD µIU/mL	CV %	SD µIU/mL	CV %
Human serum 1	0.014	0.002	11.1	0.002	11.9

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cobas e 601 and cobas e 602 analyzers						
					ermediate recision	
Sample	Mean µIU/mL	SD µIU/mL	CV %	SD µIU/mL	CV %	
Human serum 2	0.283	0.006	2.0	0.008	2.7	
Human serum 3	3.91	0.052	1.3	0.089	2.3	
Human serum 4	57.8	1.03	1.8	1.56	2.7	
Human serum 5	95.8	2.09	2.2	2.38	2.5	
PC Universal 1	1.45	0.023	1.6	0.034	2.4	
PC Universal 2	8.13	0.124	1.5	0.165	2.0	
PC Thyro Sensitive	0.184	0.004	2.1	0.005	2.9	

Method comparison

A comparison of the Elecsys TSH assay, $\boxed{\texttt{REF}}$ 08429324190 (**cobas e** 601 analyzer; y) with the Elecsys TSH assay, $\boxed{\texttt{REF}}$ 11731459122 (**cobas e** 601 analyzer; x) gave the following correlations (µIU/mL):

Number of samples measured: 134

Passing/Bablok ⁹	Linear regression
y = 1.00x + 0.004	y = 0.965x + 0.107
т = 0.966	r = 0.999

The sample concentrations were between 0.005 and 94.1 µIU/mL.

Analytical specificity

The following cross-reactivities were found, tested with a TSH concentration of approximately 0.35 $\mu\text{IU/mL}.$

Cross-reactant	Concentration tested mU/mL	Cross-reactivity %
LH	10000	0.000
FSH	10000	0.000
hGH	1000	n. d. ^{c)}
hCG	50000	n. d.

c) n. d. = not detectable

References

- Ando H. Subchapter 23C Thyroid-stimulating hormone. In: Ando H, Ukena K, Nagata S, editors. Handbook of Hormones (Second Edition). San Diego: Academic Press; 2021. p. 193-195.
- 2 Ellervik C, Halsall DJ, Nygaard B. Thyroid Disorders. In: Rifai N, Chiu RWK, Young I, Burnham CAD, Wittwer CT, editors. Tietz Textbook of Laboratory Medicine, Saunders Elsevier, Philadelphia, 7th edition, 2023, chapter 57, p. 806-845.e13.
- 3 Garber JR, Cobin RH, Gharib H, et al. American Association of Clinical Endocrinologists and American Thyroid Association Taskforce on Hypothyroidism in Adults. Clinical practice guidelines for hypothyroidism in adults: cosponsored by the American Association of Clinical Endocrinologists and the American Thyroid Association. Endocr Pract. 2012 Nov-Dec;18(6):988-1028. doi: 10.4158/EP12280.GL. Erratum in: Endocr Pract. 2013 Jan-Feb;19(1):175.
- 4 Ross DS, Burch HB, Cooper DS, et al. 2016 American Thyroid Association Guidelines for Diagnosis and Management of Hyperthyroidism and Other Causes of Thyrotoxicosis. Thyroid. 2016 Oct;26(10):1343-1421. doi: 10.1089/thy.2016.0229. Erratum in: Thyroid. 2017 Nov;27(11):1462.
- 5 Persani L, Brabant G, Dattani M, et al. 2018 European Thyroid Association (ETA) Guidelines on the Diagnosis and Management of Central Hypothyroidism. Eur Thyroid J 2018 Oct;7(5):225-237. doi: 10.1159/000491388.
- 6 Urgatz B, Razvi S. Subclinical hypothyroidism, outcomes and management guidelines: a narrative review and update of recent literature. Curr Med Res Opin 2023 Mar;39(3):351-365. doi: 10.1080/03007995.2023.2165811.

- 7 Sakai H, Fukuda G, Suzuki N, et al. Falsely Elevated Thyroid-Stimulating Hormone (TSH) Level Due to Macro-TSH. Endocr J 2009;56(3):435-440.
- 8 Ebert C, Bieglmayer C, Igari J, et al. Elecsys TSH, FT4, T4, T-uptake, FT3 and T3. Clinical results of a multicentre study. Wien Klin Wochenschr 1998;110 Suppl 3:27-40.
- 9 Bablok W, Passing H, Bender R, et al. A general regression procedure for method transformation. Application of linear regression procedures for method comparison studies in clinical chemistry, Part III. J Clin Chem Clin Biochem 1988 Nov;26(11):783-790.

For further information, please refer to the appropriate operator's manual for the analyzer concerned, the respective application sheets and the Method Sheets of all necessary components (if available in your country).

A point (period/stop) is always used in this Method Sheet as the decimal separator to mark the border between the integral and the fractional parts of a decimal numeral. Separators for thousands are not used.

Any serious incident that has occurred in relation to the device shall be reported to the manufacturer and the competent authority of the Member State in which the user and/or the patient is established.

Symbols

Roche Diagnostics uses the following symbols and signs in addition to those listed in the ISO 15223-1 standard (for USA: see dialog.roche.com for definition of symbols used):

CONTENT	Contents of kit
SYSTEM	Analyzers/Instruments on which reagents can be used
REAGENT	Reagent
CALIBRATOR	Calibrator
\longrightarrow	Volume for reconstitution
GTIN	Global Trade Item Number

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